

A Systematic Review on Molluscicidal Activity of Plants Worldwide Against Intermediate Snail Host of Schistosomes

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Article Info	ABSTRACT
<p>Article type: Review Article</p> <p>Article History: Received: 4 Feb 2025 Revised: 9 May 2025 Accepted: 31 May 2025 Published Online:</p> <p>✉ Correspondence to: Naseer Ali Shah</p> <p>Email: drnaseer@comsats.edu.pk</p>	<p>Objective: Schistosomiasis affects more than 230 million people worldwide each year. Aquatic snails, which serve as the intermediate hosts, play a key role in transmitting the disease. The present systematic review compiles data on the molluscicidal activity of plants and their phytochemicals to help minimize the incidence of schistosomiasis.</p> <p>Methodology: Online databases such as Google Scholar, PubMed, and ScienceDirect were searched for articles published between 2000 and 2022. In total, 290 articles were identified, of which 193 were included; studies involving snail species not associated with schistosomiasis were excluded. Data such as snail species, plant species and family name, plant parts used, assessment type, and phytoconstituents were analyzed.</p> <p>Results: This review identified 279 plant species with molluscicidal activity, most belonging to the families Euphorbiaceae (9.31%), Solanaceae (7.16%), Asteraceae (6.09%), Leguminosae (5.73%), and Lamiaceae (4.3%). The most frequently used plant parts were leaves (48.74%), followed by seeds (12.9%), fruits (15.05%), roots (14.69%), stems (12.54%), and bark (10%).</p> <p>Conclusion: Moreover, the review highlights the major plant phytochemicals and mechanisms of molluscicidal activity. Plants and their phytochemicals are promising resources for controlling snail species associated with schistosomiasis. Future studies should focus on identifying and isolating active phytochemicals and clarifying their mechanisms of molluscicidal action. Keywords: Schistosomiasis Control, Phytochemicals, Molluscicidal Plants, Biomphalaria, Oncomelania, Intermediate Host</p>
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Introduction

Schistosomiasis, commonly known as snail fever, is caused by trematodes belonging to the genus *Schistosoma* [1]. The main *Schistosoma* species that cause schistosomiasis include *S. japonicum*, *S. mansoni*, *S. haematobium*, *S. mekongi*, and *S. intercalatum* [2]. Schistosomiasis is a parasitic disease caused by *Schistosoma* worms, with clinical manifestations varying depending on the species, immune response, and infection duration. Symptoms include fever, fatigue, eosinophilia, abdominal pain, rashes, cough, respiratory issues, muscle and joint pain, nausea, vomiting, and gastrointestinal disturbances. Chronic symptoms include hepatosplenomegaly, intestinal complications, urinary symptoms, neurological symptoms, immunoreconstitution inflammatory syndrome, and cognitive

impairments. Understanding these symptoms can aid in early detection and management, reducing the burden of schistosomiasis on affected individuals and communities [3].

Globally, over 230 million people are infected, with Africa accounting for 95% of cases, highlighting significant regional disparities in disease burden [4]. The regional breakdown of the numbers shows that Africa has the highest prevalence due to a combination of environmental and socioeconomic factors that favour parasite survival. In Uganda, [5] even after treatment, individuals in high-endemic areas continue to become infected. Schistosomiasis in Asia varies by region, but rural populations in China show a high prevalence [6].

The life cycle of schistosomes is completed in two hosts. Snails, mainly freshwater snails, serve as an intermediate host in which schistosomes undergo asexual reproduction, including the development of miracidia and formation of cercariae (a tailed structure), which upon contact with human skin enter the tissue and develop into adults [1]. The snail species acting as an intermediate host of schistosomes [7] mainly belong to the genera *Biomphalaria*, *Bulinus* and *Oncomelania* (Gastropoda: Pulmonata: Planorbidae) [8,9].

Biomphalaria glabrata, host of *S. mansoni*, is widely found in Africa and South America and sustains transmission in both urban and rural settings. In East and Southeast Asia, *Oncomelania hupensis* is closely linked with *S. japonicum*, particularly in rice paddies where irrigation practices increase exposure risks. *Bulinus truncatus*, the main host of *S. haematobium*, thrives in freshwater habitats across the Middle East and North Africa, playing a central role in urinary schistosomiasis. In contrast, *Austropeplea* species have a more restricted distribution in Sub-Saharan Africa and mainly facilitate livestock-associated *S. bovis* infections. Together, these host–parasite associations underline the ecological diversity of transmission and can help novice readers appreciate regional variations in schistosomiasis epidemiology [10].

In different regions of the world, various control strategies are used to prevent the transmission and outbreak of schistosomiasis [11] and eliminate the disease by disrupting the life cycle of snail species (intermediate host) [12]. Egypt was the first country to control schistosomiasis by using chemical agents against the schistosome-transmitting snail species [13]. The chemicals commonly used as molluscicides include pyrethroids, carbamates, metaldehyde, organophosphates, and niclosamide [14]. Metaldehyde is a widely used, stable molluscicide belonging to the aldehyde group; its application causes snails to secrete an excessive amount of mucus from the body, leading to dehydration and death [15]. Niclosamide is another preferred synthetic molluscicide that restricts the transmission of schistosomes by killing snails or making them inactive [16]. However, these synthetic molluscicides cause environmental pollution.

Recently, the focus has shifted from synthetic molluscicides to plant-based molluscicides because they are economical, easily degradable in the environment and less likely to produce resistance in snails [17]. *Phytolacca dedecandra* (family Phytolaccaceae) is the first plant reported to exhibit molluscicidal activity [18], but its use is now restricted owing to its toxicity to non-target species [11].

An ideal molluscicidal plant must have certain key characteristics. For example, the plant extract should be toxic at extremely low concentrations and non-toxic to non-target species and it needs to remain viable after a period of storage. It is also preferred that the plant species is endemic. Another characteristic of an ideal plant-based molluscicide is that it has a stable structure against environmental fluctuations [19].

This review aims to present worldwide data on plants and their phytochemicals that exhibit molluscicidal activity against intermediate host snail species of schistosomes.

Materials and Methods

Studies regarding the molluscicidal activity of plants against the intermediate host of schistosomes were identified and screened on the basis of certain inclusion and exclusion criteria described later here (Fig 2). Articles were searched using online databases such as Google Scholar, PubMed, and ScienceDirect using keywords such as “plant molluscicide,” “phytolacca, natural products against schistosome snail hosts.phytochemicals,” “molluscicidal activity of plants,” “snail sp. causing schistosomiasis” and “plant extracts against snail species.” Only articles available as full texts (n = 290) were downloaded and further screened; of these, duplicates (n = 6), irrelevant studies (n = 4), and studies that did not mention snail species associated with the transmission of schistosomiasis (n = 83) were excluded. The remaining 193 articles were included in this review. Subsequently, data were extracted from 173 articles (Table 1), including information on plant species and family, plant parts used, screened phytochemicals, and their efficacy against snail species.

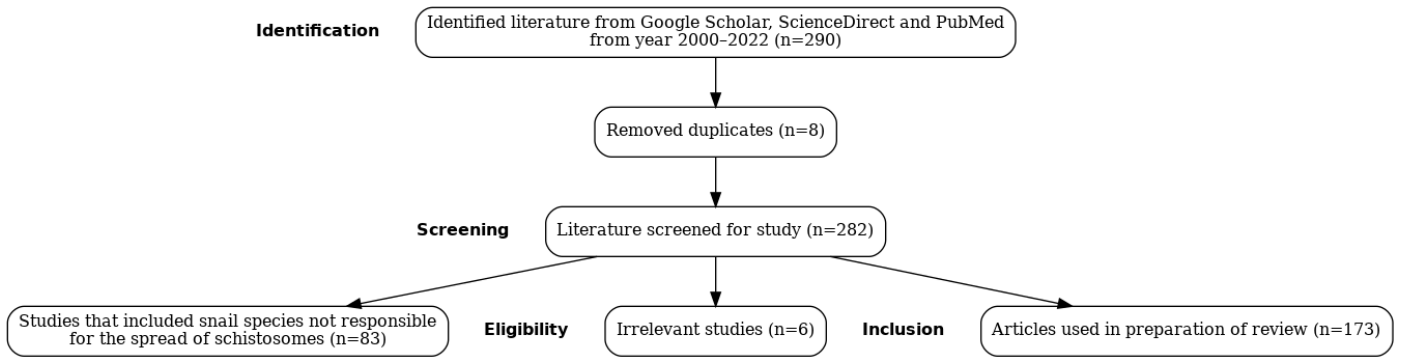


Figure 1: Study design for the review process.

Results

A total of 193 articles were found eligible to be included in the review, and data from 173 articles related to plant molluscicidal activity against schistosome-transmitting snails were extracted (Table 1).

Table 1: List of plants exhibiting molluscicidal activity against snail species that acts as intermediate hosts of schistosomes (data extracted from articles published between 2000 and 2022)

Snail species	Country	Plant species	Plant Family	Part used	Extraction method	Phytoc constituent	Assessment type	Lethal concentration	Findings	Ref.
Biomophalaria alexandrina	Egypt	Moringa oleifera	Moringaceae	Seed	Cerium oxide nano-composite	-----	In-vitro	LC50= 314.5 mg/L, LC90= 386.5 mg/L	Reduced reproductive rate, significant disappearance of cilia and structural deformation in tubular glands	[20]
	Egypt	Colchicum ritchii	Colchicaceae	Flower	Methanolic extract	Tannins, flavonoids, saponins, terpenoids, steroids, anthraquinones and alkaloids	In-vitro, In-vivo	LC50 = 35.1 mg/L	Significant increase in micronuclei and 55 % miracidia mortality rate	[21]
	Egypt	Ziziphus spina-christi	Rhamnaceae	Seed	Methanolic extract	Tannins, saponins, alkaloids, flavonoids, phenols and glycosides	In-vitro	LC50 = 108 mg/L	Reduced catalase activity, Increased total antioxidant activity and caused mortality in Daphnia magna	[22]

	Egypt	<i>Carica papaya</i>	Caricaceae	Seed	Methanolic extract	Tannin and saponin	In-vitro	LC50= 138.5 mg/L	Increased catalase activity, Slight increase in total antioxidant activity and caused mortality in <i>Daphnia magna</i>	
	Egypt	<i>Origanum majorana</i>	Lamiaceae	Leaves	Aqueous extract	Zingiberene, nerolidol, cubenol, eudesmenol, cedren, butanone, capsaicin, gingerol, stigmasterol and nonivamide	In-vitro	LC50= 42 mg/ml	Swelling of a cephalopodal mass and hemorrhagic blisters on the foot sole	[23]
	Egypt	<i>Ziziphus christi</i> spina-	Rhamnaceae	Leaves	Aqueous extract	Cyclooctasiloxane, hydrochinon, morphinan, tricycloundecan, spathulenol, imidazole, megastigmatrieno	In-vitro	LC50= 43 mg/ml	Reduced miracidial activity and 100 % cercarial activity at 6.25 mg/ml	[23][22][22][22][22][22][22]

						ne, guanosin e, desulpho sinigrin, phenanth renemeth anol and thienopyr idine				
	Egypt	Salvia fruticosa	Lamiaceae	Leaves	Aqueous extract	Quinol ol, tetrasilox ane, androstad ienol, purinol, spirosten, tetradeca noic acid, benzene methanol and cystathio nineand terphenyl	In-vitro	LC50= 69 mg/ml	Less effective against miracidial activity and 100 % cercarial activity at 100 mg/ml	
	Egypt	Ziziphus christi spina	Rhamnaceae	Leaves	Ethanollic extract	-----	In-vitro	LC50=	Increased apoptosis and condensed zygotene	[24]

								108.7 mg/l		
Egypt	<i>Moringa oleifera</i>	Moringaceae	Leaves	Ethanollic extract	-----	In-vitro	LC50= 209.4 mg/l	Increased apoptosis and condensed zygotene		
Egypt	<i>Tecoma stans</i>	Bignoniaceae	Leaves	Ethanollic extract	-----	In-vitro	LC50= 256 mg/l	Increased apoptosis and condensed zygotene		
Egypt	<i>Carica papaya</i>	Caricaceae	Leaves	Methanol, ethanol and butanol extract	----- -	In-vitro	LC50 (mg/L) against cercariae For methanol= 2; For ethanol= 20 and for butanol= 4	65-86 % mortality rate	[25]	
Egypt	<i>Anagallis arvensis</i>	Primulaceae	Whole plant	Ethanollic extract	----- --	In-vitro	LC50= 1.76 ppm ; LC90= 3.16 ppm	Reduction in protein content, denatured hermaphrodite gland, decreased amount of connective tissues and 100 % mortality rate in case of LC90 after 4 hr	[26]	
Egypt	<i>Anagallis arvensis</i>	Agavaceae	Whole plant	Methanolic extract	----- ---	In-vitro	LC50= 45 ppm LC90= 60 ppm	Extensive vacuolation in cells of digestive system and 100 % mortality at 70 ppm	[27]	
Egypt	<i>Viburnum tinus</i>	Caprifoliaceae	Whole plant	Methanolic extract	----- ---	In-vitro	LC50= 38 ppm	90 % mortality at 60 ppm, deformed gonadal cells, reduced		

								LC90= 59 ppm	number of sperms and degenerated cercariae	
Egypt	Anagallis arvensis	Primulaceae	Leaves	Aqueous extract	----- -	In-vitro		LC50= 37.9 mg/l LC90= 48.3 mg/l	Decreased reproductive rate, deformation in hermaphrodite gland cells and loss of connective tissues in gonadal cells	[28]
Egypt	Albizia anthelmintica	Leguminosae	Leaves	-----	Saponin	In-vitro		LC50 = 17.6 ppm LC90 = 25.3 ppm	Decrease in granulocytes and hyalinocytes while increased amoebocytes	[29]
Egypt	Callistemon citrinus	Myrtaceae	Leaves	Methanol extract	----- -	In-vitro		LC90= 37.8 ppm	Loss of hyaline substance, loss of connective tissues and formation of vacuoles	[30]
Egypt	Callistemon citrinus	Myrtaceae	Leaves	Methanolic extract	----- -	In-vitro		LC50= 22.3 ppm LC90 = 37.8 ppm	54.2 % reduction in reproductive rate, crumpling of young oocytes and 56% reduction of testosterone	[31]
Egypt	Zingiber officinale	Zingiberaceae	Root	Methanolic extract	----- -	In-vitro		LC50= 43.8 ppm LC90 = 74.9 ppm	60 % reduction in reproductive rate, atrophy of nuclei and degeneration of necrotic cells	

Egypt	<i>Asparagus densiflorus</i>	Asparagaceae	Leaves	Aqueous extract	----- -	In-vivo	LC50 = 75 ppm LC90 = 102 ppm	Caused atrophy in gonadal cells, reduced reproductive rate and 100% mortality	[32]
Egypt	<i>Oreopanax guatemalensis</i>	Araliaceae	Leaves	Aqueous extract	----- -	In-vivo	LC50 = 160 ppm LC90 = 188 ppm	100 % motality rate at 1000 mg/L within 24 h	
Egypt	<i>Juniperus horizontalis</i>	Cupressaceae	Aerial parts	Methanolic extract	Thujone, D-limonene, β -myrcene, sabinene and methyl citronella te	In-vitro	LC50 = 38.9 ppm	Proved toxic against snail sp.	[33]
Egypt	<i>Juniperus communis</i>	Cupressaceae	Aerial parts	Methanolic extract	Sabinene, β -myrcene, D-limonene,	In-vitro	LC50 = 22.9 ppm	Proved toxic against snail sp.	

						thujone and terpinen-4-ol				
	Egypt	Cestrum purpureum	Solanaceae	Whole plant	Methanolic extract	ursolic acid and 3-O-β-D-glucopyranosyl-(1→4)-α-L-rhamnopyranosyl-2 and 19-dihydroxy-epihederagenin-28-O-1-[26-(3-acetyl-4-methoxy-cinnamoyloxyhexacosanoyl)] glycerol	In-vivo	LC50= 72 ppm LC90 = 120 ppm	90 % mortality rate after 24h	[34]
	Egypt	Agave angustifolia	Agavaceae	Whole plant	Aqueous suspension	----- -	In-vivo, In-vitro	LC90= 120 ppm	Decreased egg laying capacity and survival rate	[35]

Egypt	<i>Pittosporum tobira</i>	Pittosporaceae	Whole plant	Aqueous suspension	----- -	In-vivo, In-vitro	LC90= 120 ppm	Decrease in cercaricidal mortality by 81% and miracidia by 87 %	
Egypt	<i>Euphorbia peploides</i>	Euphorbiaceae	Aerial parts	Methanolic extract	Quercetin, luteolin, gallic acid, β -amyryn, P-hydroxybenzoic acid, quercetin-3-O- β -D-glucopyranoside and kampferol-3-O- β -D-glucopyranoside	In-vivo	LC90= 30 ppm	Strong molluscicidal activity	[36]
Egypt	<i>Callistemon viminalis</i>	Myrtaceae	Leaves, fruit and bark	Methanolic extract	----- -	In-vivo	LC50 (ppm): Fruit=6.2, Bark=32, Leaves =40	63.3 % mortality rate due to leaves extract, 66.7% due to bark and 100% due to leaves	[37]
Egypt	<i>Eucalyptus globulus</i> , <i>Melaleuca styphelioides</i>	Myrtaceae	Leaves	Methanolic extract	Eucalbanin B, tellimagrandin I, pentagal	In-vitro	LC50= 165.6 ppm	25 % reduction in protein content, 100 % cercaricidal mortality, 21 % increase in acid	[38]

						oyl glucose, casuarini n and 1,2,3,4,6- Penta-O- galloyl-b- D- glucopyr anose		LC90= 245.5 ppm	phosphatase and 113 % increase in aspartate aminotransferase	
	Egypt	Citrus limon	Rutaceae	Peel	Aqueous extract	----- --	In-vitro	LC50 =744.17 ppm LC90= 1821.25 ppm	58 % reduction in hatchability rate, necrosis in digestive glands and significant increase in protein content	[39]
	Egypt	Origanum syriacum	Lamiaceae	Seed	Ethanollic extract	----- --	In-vitro	B. alexandrina LC50= 226.8 ppm LC90= 422.9 ppm	Chromosomal aberrations	[40]
	Egypt	Yucca desmettiana	Asparagaceae	Leaves	Methanolic extract	Saponin	In-vivo	LC50 = 68 mg/L LC90 = 96 mg/L	Significant molluscicidal activity	[41]

Egypt	<i>Haptophyllum tuberculatum</i>	Rutaceae	Aerial parts	Chloroform and methanolic extracts	Steroids	In-vitro	LC50 = 16 ppm	100 % mortality rate at 25 ppm with excessive mucous production, increased hemolysis and decreased progesterone level	[42]
Egypt	<i>Datura stramonium</i>	Solanaceae	Leaves	Dry powder	----- -	In-vitro	LC50= 25.8 ppm LC90= 40.4 ppm	60 % mortality rate, increased alanine transaminase and alkaline phosphatase in haemolymph	[43]
Egypt	<i>Sesbania sesban</i>	Leguminosae	Leaves	Dry powder	----- -	In-vitro	LC50= 51.0 ppm LC90= 62.4 ppm	80 % mortality rate and decreased total protein content	
Egypt	<i>Solanum sinicum</i>	Solanaceae	Leaves	Ethanollic, aqueous, methanolic and chloroform extract	----- -	In-vitro	LC50 = 14.8 ppm	88 % reduction in reproductive rate, 100 % increase in glucose and 80 % in lactate and significant decrease in glycogen, pyruvate and total protein amount	[44]
Egypt	<i>Artemisia judaica</i> L	Asteraceae	Leaves	Ethanollic, aqueous, methanolic and chloroform extract	----- -	In-vitro	LC50 = 38 ppm	Increase in protease quantity but decreased lactic dehydrogenase and cytochrome oxidase	
Egypt	<i>Dalbergia sissoo</i>	Leguminosae	Fruits, leaves, root and stem bark	Ethanollic extracts	----- -	In-vivo	LC50= 10.8 ppm LC90= 20.4 ppm	Mainly toxic at pre-hatch stage and 100 % mortality rate	[45]

	Egypt	<i>Furcraea selloa marginata</i>	Agavaceae	Leaves	Aqueous suspension	----- -	In-vitro	LC50 =53.66 ppm LC90 =84.35 ppm	30 % mortality rate after 24 hr and 100 % miracidial mortality rate	[46]
	Egypt	<i>Euphorbia milii</i>	Euphorbiaceae	Latex	Aqueous solution	----- -	In-vitro	LC50= 19 ppm LC90= 38 ppm	Proved toxic against snail sp.	[47]
	Egypt	<i>Meryta denhamii</i>	Araliaceae	Flower	Methanolic extract	Oleonolic acid-3-O- α -arabinopyranoside, oleonic acid-3-O- β -glucopyranoside and hedragenin-3-O- α -rhamnopyranoside	In-vivo	LC50=85 mg/L	Increased glucose level and decreased glycogen and protein level in tissues	[48]
	Egypt	<i>Guayacum officinalis</i>	Zygophyllaceae	Whole plant	Methanolic, aqueous and ethanolic extract	----- -	In-vitro	LC50= 120 ppm LC90= 210 ppm	Decreased acetylcholinesterase and Succinate dehydrogenase	[49]

Egypt	<i>Atriplex stylosa</i>	Chenopodiaceae	Whole plant	Methanolic, aqueous and ethanolic extract	----- -	In-vitro	LC50= 94 ppm LC90= 180 ppm	36 % decreased glycogen content and 53 % increased glucose level	
Egypt	<i>Euphorbia splendens</i>	Euphorbiaceae	Whole plant	Methanolic, aqueous and ethanolic extract	-----	In-vitro	LC50= 40 ppm LC90= 73 ppm	Effectively decreased the number of juveniles and adult	
Egypt	<i>Citrus reticulata</i>	Rutaceae	Fruit	Peel extract	-----	In-vivo	LC50= 163.79 ppm	95 % mortality rate	[50]
Egypt	<i>Euphorbia paralias</i>	Euphorbiaceae	Aerial parts	Methanolic extract	Diterpene (Paralialanes, segetanes and jatrophenes)	In-vivo	-----	100 % mortality rate	[51]
Egypt	<i>Commiphora molmol</i>	Bruseraceae	Stem (Gum resin)	Aqueous solution	Terpenes and eugenol	In-vivo	-----	67 % mortality rate	[52]
Egypt	<i>Commiphora molmol</i>	Bruseraceae	Whole plant	Oil extract	----- -	In-vivo	LD50= 155 ppm LD90= 195 ppm	100 % mortality rate	[53]

	Egypt	<i>Yucca elephantipes</i>	Agavaceae	Leaves	Methanolic extract	saponin	In-vivo	LC90= 4 ppm	Strong molluscicidal activity	[54]
	Egypt	<i>Ambrosia Maritima</i>	Asteraceae	Whole plant	Dry powder	----- ---	In-vivo	-----	95 % decrease number of snails in first month	[55]
	Egypt	<i>Azolla pinnata</i>	Salviniaceae	Whole plant	Dry powder	----- ---	In-vivo	-----	94 % decrease number of snails in first month	
	Egypt	<i>Commiphora molmol</i>	Burseraceae	Whole plant	Oil and resin extract	----- ---	In-vivo	LC50= 4 ppm	100 % mortality rate of both snail sp.	[56]
	Saudi Arabia	<i>Rosmarinus officinalis</i>	Lamiaceae	Whole plant	Pet-ether and ethanolic extracts	----- -	In-vivo	LC50= 236.81 ppm	85 % mortality rate	[57]
	Italy	<i>Vitex trifolia</i>	Verbenaceae	Leaves, stem and seed	chloroform, n-hexane and ethanolic extract	Platonic acid , β -sitosterol, ursolic acid acetate and ursolic acid acetate	In-vivo	LC50= 26.42 mg/l	92 % mortality rate at 32.01 mg/l	[58]
<i>Biomphalaria pfeifferi</i>	Zimbabwe	<i>Cucurbita maxima</i>	Cucurbitaceae	Seed	Aqueous and crude ethanolic extracts	----- -	In-vivo	LC50 (mg/ml) for:	100 % mortality rate at 2 mg/mL in both snails	[59]

								Aqueous extract = 0.002, Ethanollic extract = 0.002		
	Nigeria	Balanites aegyptiaca	Balanitaceae	Leaves and fruit	Aqueous extract	Alkaloids, saponins, flavonoids, tannins, phenols and steroids	In-vitro	-----	90 % mortality rate at 50 µg/ml	[60]
	Sudan	Combretum glutinosum Perr. ex DC	Combretaceae	leaves	Ethanollic extract	Cumarins, tannins, flavonoids, saponins and triterpenes	In-vivo	LC50 at 24 hr: 117.57 LC90 at 24 hr: 220.84	Possess strong molluscicidal ability	[61]
	Sudan	Hyphaene thebaica (L.) Mart	Arecaceae	Male	Ethanollic extract	Cumarins,	In-vivo	LC50 at 24 hr:	Caused mortality at low concentration	

				Inflorescences		tannins and saponins		158 LC90 at 24 hr: 228	
Sudan	Indigofera oblongifolia Forssk	Leguminosae	Aerial parts	Ethanollic extract	Cumarins , tannins, saponins, triterpenes and steroids	In-vivo	LC50 at 24 hr: 158.11 LC90 at 24 hr: 228.11	Mortality rate was more than 90 %	
Sudan	Rhynchosia minima (L.) DC	Leguminosae	Aerial parts	Ethanollic extract	Cumarins , tannins, flavonoids and saponins	In-vivo	LC50 at 24 hr: 158.11 LC90 at 24 hr: 228.11	Caused mortality at low concentration	
Sudan	Solanum dubium L.	Solanaceae	Fruit	Ethanollic extract	Cumarins , tannins, flavonoids,	In-vivo	LC50 at 24 hr: 153.02 LC90 at 24 hr:	90 % mortality at low concentration	

						saponins, alkaloids and steroids		226.62		
	Sudan	<i>Pulicaria crispa</i>	Asteraceae	Leaves, stem and root	Powder	----- -	In-vivo	LC50(ppm): Leaves= 616.6 Stem=616.6 Root= 645.65 LC90(ppm): Leaves= 776.25 Stem= 812.83 Root= 812.83	Sluggish and disoriented behavior	[62]
	Ethiopia	<i>Achyranthes aspera</i>	Amaranthaceae	Leaves	Hydro-ethanolic extract	Flavonoids, saponins, alkaloids and terpenoids	In-vivo	In immersion: LC50(ppm) = 20.37 LC90 (ppm) = 46.84; In immersion:	30-40 % mortality rate after 24 hr at 100 ppm	[63]

								LC50 = 3.10 ppm LC90 = 11.08 ppm		
	Sudan	Solenostemma argel	Asclepiadaceae	Leaves	Aqueous extract	----- -	In-vivo	For egg masses, LD50 = 2921.3 ppm LD95 = 3515 ppm; For neonates, LD50 = 368 ppm, LD95 = 1699 ppm	50 % mortality rate of eggs and 95 % of neonates	[64]
	Kenya	Aloe secundiflora	Asphodelaceae	Leaves	Aqueous extract	----- -	In-vivo	-----	40 % mortality rate after 24 h	[65]
	Kenya	Balanites aegyptiaca	Zygophyllaceae	Root	Aqueous extract	----- -	In-vivo	-----	36 % mortality rate after 24 h	
	Kenya	Aspilia pluriseta	Asteraceae	Leaves	Aqueous extract	----- -	In-vivo	-----	50 % mortality rate after 24 h	
	Kenya	Psidium guajava	Myrtaceae	Root and leaves	Aqueous extract	----- -	In-vivo	-----	20 % mortality rate after 24 h	

	Kenya	<i>Azadirachta indica</i>	Meliaceae	Root and leaves	Aqueous extract	----- -	In-vivo	-----	58 % mortality rate after 24 h	
	Sudan	<i>Solenostemma argel</i>	Apocynaceae	Leaves	Aqueous extract	Alkaloids , terpenoids, flavonoids, saponins and tannins	In-vivo	LD50= 0.103 ppm LD95= 0.187 ppm	Behavioral changes	[66]
	Nigeria	<i>Balanites aegyptiaca</i>	Zygophyllaceae	Stem and bark	Aqueous solution	2-methoxy-4-vinylphenol, benzaldehyde, phenol, 2-methoxy-4-(2-propenyl)-acetate, ethanone, 1-(4hydroxyphenyl)-, alpha-d-6,3-furanose, 4-(3-hydroxy-	In-vivo	LD50 (mg/L): For B. pfeifferi juvenile=1.4 For B. pfeifferi adult=1.6 LD90 (mg/L): For B. pfeifferi juvenile=4.8 For B. pfeifferi adult=4.2	100 % mortality rate of both snail species at 40mg/L	[67]

						1-propenyl) -2-methoxy phenol, hexadecanoic acid, 2-butanone, 9-octadecanoic acid, octadecanoic acid, 13-octadecenal, tetradecanoic, 9-octadecanamide and di-n-octylphthalate				
	Mali	<i>Glinus oppositifolius</i>	Aizoaceae	Whole plant	Crude extract	12-Glucosyl-GO1	In-vivo	LC50 of Butanol extract= 91.7 ppm; LC50 of Ethanolic extract= 116.6 ppm	100 % mortality rate of both snail species at 200ppm	[68]

Ethopia	<i>Glinus lotoides</i>	Molluginaceae	Fruit	Aqueous and crude extract	----- -	In-vitro	LC50= 47.1 mg/L LC90= 56.96 mg/L	20 % mortality rate of cercariae, and 100 % snail mortality at 37.5-80 mg/L	[69]
Ethopia	<i>Balanites aegyptiaca</i>	Zygophyllaceae	Seeds, mesocarp, fruit and endocarp	Aqueous extract	----- -	In-vivo	LC90 (mg/L): Seed= 77.70, Endocarp= 120.04, Mesocarp= 89.50, Fruit=99.55	90 % mortality rate	[70]
Kenya	<i>Entada leptostachya</i>	Leguminosae	Root	Ethyl acetate, aqueous and methanol extract	Tannins, glycosides, Saponins, flavonoids and triterpenes	In-vitro	LD50 (mg/l): Juvenile= 30.12 Adult= 40.93	100 % mortality rate on juveniles and 95 % on adult	[71]
Kenya	<i>Azadirachta indica</i>	Meliaceae	Leaves	Ethyl acetate, aqueous, methanol extract	Alkaloids, glycosides, saponins, flavonoids and	In-vitro	-----	Only 5 % mortality	

						triterpenes				
	Nigeria	Chromolaena odorata	Asteraceae	Leaves	Aqueous and ethanolic extract	----- --	In-vivo	LC50 (ppm) for Aqueous extract: Eggs= 65.75 Juveniles= 75.59 Adults= 217.57 LC50 (ppm) for Ethanolic extract: Eggs= 44.03 Juveniles= 44.68 Adults= 88.04	100 % mortality rate after 24 h	[72]
	Nigeria	Ficus exasperata	Moraceae	Root, bark, seed and leaves	Aqueous and ethanolic extracts	Flavonoids, alkaloids, tannins, saponins, anthraquinones and cardenolides	In-vitro	LC50= 0.3187 ppm LC90= 0.7555 ppm	Mitochondrial eruption, degeneration of epithelial cells and 100 % mortality rate	[73]

	Nigeria	<i>Cymbopogon citratus</i>	Poaceae	Leaves	Ethanol and aqueous extract	----- -	In-vivo	LC50 for aqueous extract= 140.74 ppm LC50 for ethanolic extract= 254.92 ppm	100 % mortality in ethanol extract	[74]
	Nigeria	<i>Chromolaena odorata</i>	Asteraceae	Leaves	Ethanol and aqueous extract	----- -	In-vivo	Aqueous extract: LC50 =65.75 ppm LC90=139.54 ppm Ethanolic extract LC50 =44.03 ppm LC90 =119.3 ppm	100 % mortality rate at 100 ppm	[75]
	Sudan	<i>Acacia seyal</i>	Mimosaceae	Bark	Methanolic extract	----- -	In-vitro	IC50 (ppm): At 24 h =80.79, At 48 h =34.33	100 % mortality at 62.5 ppm after 48 h	[76]

	Nigeria	<i>Piper guineense</i>	Piperaceae	Fruit	Aqueous and ethanolic extract	----- -	In-vivo	For ethanolic extract: LC50 = 0.10 mg/L LC90= 0.9 mg/L For aqueous extract: LC50 = 5.0 mg/L LC90= 8.5 mg/L	90% mortality rate at 9 mg/L	[77]
	Nigeria	<i>Anonna senegalensis</i>	Annonaceae	Leaves and root	Ethanolic extract	----- -	In-vivo	For leaves: LC50 = 88.93 mg/l LC90= 285.44 mg/l For roots: LC50 = 861.38 mg/l LC90= 2379.80 mg/l	100 % mortality rate of adult snail at 500 mg/l and 75% death rate of egg masses at 250 mg/l	[78]
	Nigeria	<i>Anogeissus leiocarpus</i>	Combretaceae	Leaves and	Ethanolic extract	----- -	In-vivo	For leaves:	65 % mortality rate of adult snail at 1000 mg/l and 90% death rate of egg masses at 4000 mg/l	

				root				LC50 = 1 213.39 mg/l LC90= 3 263.00 mg/l For roots: LC50 = 653.60 mg/l LC90= 1 503.03 mg/l	
	Nigeria	<i>Crotalaria retusa</i>	Leguminosae	Leaves and root	Ethanollic extract	----- -	In-vivo	For leaves: LC50 = 73.84 mg/l LC90= 176.94 mg/l For roots: LC50 = 290.35 mg/l LC90= 714.89 mg/l	70 % mortality rate of adult snail at 500 mg/l, 69 % death rate of egg masses at 1000 mg/l
	Nigeria	<i>Dalbergia sissoo</i>	Leguminosae	Fruit, stem and bark	Ethanollic extract	----- -	In-vivo	For fruits: LC50 = 30.74 mg/l LC90= 74.33 mg/l For bark:	85 % mortality rate of adult snail at 500 mg/l and 81 % death rate of egg masses at 500 mg/l

								LC50 = 399.86 mg/l LC90= 114.84 mg/l		
	Nigeria	Vernonia amygdalina	Asteraceae	Leaves and root	Ethanollic extract	----- -	In-vivo	For leaves: LC50 = 1913.45mg/l LC90= 6241.61 mg/l For roots: LC50 = 917.79 mg/l LC90= 2384.78 mg/l	85 % mortality rate of adult snail at 2000 mg/l and 89 % death rate of egg masses at 4000 mg/l	
	Nigeria	Carica papaya	Caricaceae	Leaves	Ethanollic extract	----- -	In-vivo	LC50 B. pfeifferi=2716.3 ppm LC90= 4515.9 ppm	50 % mortality rate	[79]
	Nigeria	Terminalia catappa	Combretaceae	Leaves	Ethanollic extract	----- -	In-vivo	LC50 = 864.1 ppm LC90= 1222.8 ppm	100 % mortality rate	

Sudan	<i>Cymbopogon nervatus</i>	Poaceae	Leaves	Essential oils	----- -	In-vivo	LD50 = 213.099 ppm	100 % mortality rate at 400 ppm	[80]
Sudan	<i>Boswellia papyrifera</i>	Burseraceae	Leaves	Essential oils	----- -	In-vivo	LD50 213.31 ppm	90 % mortality rate at 350 ppm	
Nigeria	<i>Dalbergia sissoo</i>	Leguminosae	Fruit, root, stem and leaves	Ethanollic and aqueous extracts	----- -	In-vitro	LC90 =89.29 mg/l	93 % death rate of snail eggs at 500 mg/l	[81]
Nigeria	<i>Dalbergia sissoo</i>	Leguminosae	Fruit, root, leaves and stem	Aqueous and ethanolic extract	----- -	In-vitro	-----	Un-hatched embryo death	[82]
South Africa	<i>Barringtonia racemosa</i>	Lecythidaceae	Fruit and seed	Aqueous extract	----- -	In-vitro	LD50 for fruit= 521.48 ppm LD50 for seed= 556.23 ppm	More toxic effects are produced by fruit extract	[83]
Sudan	<i>Azadirachta indica</i>	Meliaceae	Leaves and seeds	Aqueous extract	----- -	In-vivo	-----	75 % decrease in snail species after 5 days	[84]
Mali	<i>Cussonia barteri</i>	Araliaceae	Roots	Methanolic and aqueous extract	----- -	In-vitro	-----	Extract proved toxic against snail sp.	[85]
Mali	<i>Glinus oppositifolius</i>	Aizoaceae	Whole plant	Methanolic and aqueous extract	----- -	In-vitro	-----	Extract proved toxic against snail sp.	

	Mali	<i>Lannea velutina</i>	Anacardiaceae	Leaves	Methanolic and aqueous extract	----- -	In-vitro	-----	Extract proved toxic against snail sp.	
	Kenya	<i>Solanum aculeastrum</i>	Solanaceae	Root, bark and fruit	Methanolic extract	----- -	In-vivo	-----	100 % mortality rate	[86]
	Saudi Arabia	<i>Jatropha glauca</i>	Euphorbiaceae	Leaves	Chloroform extract	----- -	In-vivo	LD50= 66.7 ppm LD90= 114 ppm	Decreased efficacy with increased time	[87]
	Saudi Arabia	<i>Euphorbia helioscopia</i>	Euphorbiaceae	Leaves	Methanolic extract	----- -	In-vivo	LD50= 50.8 ppm LD90= 68.2 ppm	Decreased efficacy with increased time	
	Saudi Arabia	<i>Euphorbia schimperiana</i>	Euphorbiaceae	Stem	Methanolic extract	----- -	In-vivo	LD50= 34 ppm LD90= 41.7 ppm	100 % mortality rate	
Biomphalaria glabrata	Egypt	<i>Origanum syriacum</i>	Lamiaceae	Seed	Ethanollic extract	----- -	In-vitro	LC50= 215.2 ppm LC90= 415.7 ppm	Chromosomal aberration in both snail species	[40]
	Nigeria	<i>Lycopersicon esculentum</i>	Solanaceae	Seed	Methanolic extract	----- -	In-vivo	LC50 = 873.6 ppm	78 % mortality at 1000 µg/ml	[88]

Nigeria	<i>Blighia unijugata</i>	Sapindaceae	Stem, fruit, seed and leaves	Ethanollic extract	Saponin	In-vivo	LC50= 7.6 µg/ml	100 % mortality rate at 1000 ppm	[89]
Angola	<i>Euphorbia conspicua</i>	Euphorbiaceae	Latex	Methanolic extract	----- -	In-vivo	LC100 = 1 µg/mL	Toxic against the snail sp.	[90]
Mali	<i>Cussonia barteri</i>	Araliaceae	Roots	Methanolic and aqeous extract	----- -	In-vitro	-----	Extract proved toxic against snail sp.	[85]
Mali	<i>Glinus oppositifolius</i>	Aizoaceae	Whole plant	Methanolic and aqeous extract	----- -	In-vitro	-----	Extract proved toxic against snail sp.	
Mali	<i>Lannea velutina</i>	Anacardiaceae	Leaves	Methanolic and aqeous extract	----- -	In-vitro	-----	Extract proved toxic against snail sp.	
Thailand	<i>Solanum xanthocarpum</i>	Solanaceae	Fruit	Ethanollic crude extract	----- -	In-vivo, In-vitro	LC50= 163.85 mg/l LC90= 219.33 mg/l	Extract proved toxic against snail	[91]
Pakistan	<i>Euphorbia cauducifolia</i>	Euphorbiaceae	Latex	DMSO suspension	Deoxyphorbol Esters	In-vivo	LC50=12.3 ng/ml	Induces imbalance in nutrient and water absorption	[92]
Colombo	<i>Croton floribundus</i>	Euphorbiaceae	Leaves and bark	Ethanollic and methanolic extract	Kaurenolic acid	In-vivo	LC50=4.2 µg/mL LC90=11.5 µg/mL	High molluscicidal activity	[93]
Japan	<i>Ambrosia maritima</i>	Asteraceae	Root	Chloroform extract	Ambrosin and	In-vivo	LC50 = 0.37ppm	80 % molluscicidal activity after 72h	[94]

						Pentayne ene				
	Brazil	Parkia pendula	Leguminosae	Seed	Saline extract	----- -	In-vitro, In-vivo	LC50 (mg/mL) values: Blastula= 237.19 Gastrula= 244.62 Adult= 4.99	Proved toxic at 3.0, 4.0 and 5.0 mg/mL, decrease in defense cells and detection of nuclear damage	[95]
	Brazil	Dysphania ambrosioides (L.)	Amaranthaceae	Aerial parts	Essential oil	α -Terpinene, ascaridole, thymol, carvacrol, isoascari- dole, p-cymene, limonene and γ - terpinene	In-vitro, In-vivo	LC50= 25.2 μ g/mL	100 % mortality rate at 50 μ g/mL, non-toxic to Artemia salina larvae and Danio rerio fish	[96]
	Brazil	Persea Americana Mill.	Lauraceae	Stem and bark	Ethanollic extract	Quercetin , flavonoid , anthraqui- none	In vivo, In vitro	LC50 (ppm): For embryo =27.06	Increased embryo mortality at 144 h, 100 % mortality in adult, lethargy and shell reduction after 96 h	[97]

						heterosides, coumarins and tannins		For newly hatched= 30.6 For adult =55.55		
	Brazil	Pimenta dioica	Myrtaceae	Leaves	Essential oil	Octenol, mirceno, limonene, linalol, terpinol, chavicol and eugenol	In-vivo	LC50= 18.62 mg/L	90 % mortality rate after 72 h at 75 mg/L	[98]
	Brazil	Citrus limon	Rutaceae	Fruit peel	Essential oil	Limonene, sabinene, β -pinene, γ - terpinene, β -citral, α -citral, α -pinene and β - Bisabolene	In-vivo	-----	100 % mortality at 100 mg/L	[99]

	Brazil	<i>Manilkara subsericea</i>	Sapotaceae	Leaves	Ethanollic crude extract and ethyl acetate fraction	Myrcetin, quercetin and ursolic acid	In-vivo	LD50 (µg/mL): For Ethanollic crude extract= 118.7 For ethyl acetate fraction= 23.41	53 % mortality rate at 250 µg/mL	[100]
	Brazil	<i>Euphorbia umbellata</i>	Euphorbiaceae	Latex	Latex aqueous dilutions	Triterpenes	In-vivo	LC50=1.36 mg/L LC90 =3.69 mg/L	90 % mortality rate after 24 h,	[101]
	Brazil	<i>Anadenanthera colubrina</i>	Planorbidae	Bark	Crude extract	Terpenoids, tannins and phenols	In-vivo	LC50 For adult= 0.155 mg/mL For embryo= 0.692 mg/mL	100 % mortality of adult snail and embryo at 0.5, 1.0 mg/mL respectively	[102]
	Brazil	<i>Moringa oleifera</i> Lam.	Moringaceae	Flower	Aqueous extract	Xanthone, saponin, tannin, flavanol, polyphen	In-vivo	LC50= 2.37mg/mL	95 % mortality rate at 4 mg/mL	[103]

						ol and flavones				
Brazil	<i>Jatropha elliptica</i>	Euphorbiaceae	Root	Ethanol extract	diethyl 4-phenyl-2,6-dimethyl-3,5-pyridinedicarboxylate	In-vivo	LC90=36.43 µg/mL	100 % mortality rate of schistosomes and snail	[104]	
Brazil	<i>Schinopsis brasiliensis</i>	Anacardiaceae	Stem, bark	Hydroethanol extract	Xanthones, flavanones and tannin	In-vivo	LC90=73 µg/ml	60 % mortality rate at 50 µg/ml	[105]	
Brazil	<i>Jatropha gossypifolia</i>	Euphorbiaceae	Stem, leaves and fruit	Hydroalcoholic extract	Alkaloids, tannins, triterpenoids, saponins and steroids	In-vivo	-----	100 % mortality rate at 25 ppm, significant reduction in oviposition ability and complete repression of feeding ability	[106]	
Brazil	<i>Moringa oleifera</i>	Moringaceae	Seed	Aqueous extract	-----	In-vivo	LC50 =0.419 g/l; LC90 =1.021 g/l	Toxic for <i>Biomphalaria glabrata</i>	[107]	
Brazil	<i>Piper tuberculatum</i>	Piperaceae	Stem, leaves,	Methanolic extract	Piplartine	In-vitro	LC50 (µg/ml):	100 % mortality rate of blastula, gastrula and adult	[108]	

				fruit and root				Root= 20.28 Stem=200 Leaves= 310.27 Fruit= 126.27		
	Brazil	<i>Cymbopogon winterianus</i>	Poaceae	Leaves	Essential oil	Geraniol, citronellol, citronellal and elemol	In-vivo	LC50=54.0 mg/L, LC90=97.0 mg/L,	90 % mortality rate after 72 h	[109]
	Brazil	<i>Porophyllum ruderale</i>	Asteraceae	Flower and leaves	Essential oil	β -pineno, mircene, (E)- β -ocimeno and (Z)- β -ocimeno,	In-vivo	LC50 = 774.82ppm LC90 = 812.43ppm	Proved toxic at less than 1000 ppm	[110]
	Brazil	<i>Euphorbia milii</i>	Euphorbiaceae	Latex	Aqueous solution	----- --	In-vivo	LC50= 0.7 μ l	Decreased glycogen content, increased glucose and total protein content	[111]
	Brazil	<i>Euphorbia milii</i>	Euphorbiaceae	Latex	Aqueous solution	----- --	In-vivo	EC50 = 2040 μ g/L	Increased malformation in embryos	[112]
	Brazil	<i>Lippia gracilis</i>	Solanaceae	Leaves	Essential oil	Thymol, methyl-thymol	In-vivo	LC50 = 62.2 ppm	Proved highly lethal to snail sp.	[113]

						and p-cymene		LC90= 82.8 ppm		
Brazil	<i>Turnera ulmifolia</i>	Turneraceae	Leaves	Hydroalcoholic extract	Glycosides, flavonoids, alkaloids, tannins and steroids		In-vivo	-----	No activity against snail sp.	[114]
Brazil	<i>Solanum asperum</i>	Solanaceae	Fruit	Methanolic extract	Solanandaine and solamargine		In-vivo	LC50= 25.1 µg/mL LC90= 44.1 µg/mL	90 % mortality rate after 24h	[115]
Brazil	<i>Ocotea bracteosa</i>	Lauraceae	Stem and bark	Essential oil	Cadinene, ledene and globulol		In-vivo	LC90 = 8.3 µg/mL	Proved toxic against the snail sp.	[116]
Brazil	<i>Euphorbia splendens</i>	Euphorbiaceae	Latex	Aqueous stock solution	----- --		In-vitro	LD50 = 1 mg/l LD90 = 2.3 mg/l	Increased protein conc. and decreased glycogen content	[117]
Brazil	<i>Solanum stipulaceum</i>	Solanaceae	Stem	Aqueous extract	----- --		In-vivo	LC50= 45.2 µg/mL LC90= 56.0 µg/mL	Toxic against the snail sp.	[118]

	Brazil	<i>Solanum diamantinense</i>	Solanaceae	Aerial parts	Methanolic extract	----- --	In-vivo	LC50= 33.36 µg/mL LC90= 52.8 µg/mL	High molluscicidal activity against snail sp.	[119]
	Brazil	<i>Solanum paludosum</i>	Solanaceae	Fruit	Methanolic extract	----- --	In-vivo	LC50= 48.07 µg/mL LC90= 82.86 µg/mL	High molluscicidal activity against snail sp.	
	Brazil	<i>Stryphnodendron adstringens</i>	Mimosoideae	Leaves and bark	Ethanollic extract	Flavanoi ds	In-vivo	-----	90 % mortality rate at 50 ppm	[120]
	Brazil	<i>Stryphnodendron polyphyllum</i>	Mimosoideae	Leaves and bark	Ethanollic extract	Flavanoi ds	In-vivo	-----	70 % mortality rate of adult snail	
	Brazil	<i>Dimorphandra mollis</i>	Mimosoideae	Leaves and bark	Ethanollic extract	Flavanoi ds, coumarin s and tannins	In-vivo	-----	10 % mortality rate at 100 ppm after 48 h	
	Brazil	<i>Caryocar brasiliensis</i>	Caryocaraceae	Leaves and bark	Ethanollic extract	Hydrolys able tannins, flavanoi ds and condense d tannins	In-vivo	-----	90% mortality rate at 100 ppm	
	Brazil	<i>Eugenia dysenterica</i>	Myrtaceae	Leaves and bark	Ethanollic extract	Condense d tannins, flavanoi d	In-vivo	-----	100 % mortality rate	

						s, phenolic acid and coumarin s				
Brazil	<i>Annona crassiflora</i>	Annonaceae	Leaves	Ethanollic extract	Flavanoi ds and tannin	In-vivo	-----	Less than 10 % mortality rate		
Puerto Rico	<i>Didymopanax morototoni</i>	Araliaceae	Leaves	Methanolic extract	----- --	In-vivo	-----	100 % mortality rate at 50 ppm	[121]	
Puerto Rico	<i>Mammea americana</i>	Clusiaceae	Leaves	Methanolic extract	----- --	In-vivo	-----	100 % mortality rate		
Puerto Rico	<i>Furcraea tuberosa</i>	Agavaceae	Leaves	Methanolic extract	----- --	In-vivo	-----	100 % mortality rate		
Puerto Rico	<i>Argemone mexicana</i>	Papaveraceae	Leaves	Methanolic extract	----- --	In-vivo	-----	67 % mortality rate at 100 ppm		
Puerto Rico	<i>Paullinia pinnata</i>	Sapindaceae	Leaves	Methanolic extract	----- --	In-vivo	-----	50 % mortality rate at 50 ppm		
Puerto Rico	<i>Solanum americanum</i>	Solanaceae	Leaves	Methanolic extract	----- --	In-vivo	-----	33 % mortality rate at 50 ppm		
Brazil	<i>Annona crassiflora</i>	Annonaceae	Pulp, seed, stem and root	Ethanollic extract	----- --	In-vivo	LD90 (ppm): For adult= 2.34 For egg= 1	80 % mortality rate	[122]	

Brazil	<i>Annona glabra</i>	Annonaceae	Leaves and seed	Ethanollic extract	----- --	In-vivo	LD90 (ppm): For adult= 3.79 For egg= 1	80 % mortality rate
Brazil	<i>Annona muricata</i>	Annonaceae	Leaves and stem bark	Ethanollic extract	----- --	In-vivo	LD90 (ppm): For adult= 3.79 For egg= 1	60 % mortality rate
Brazil	<i>Annona pisonis</i>	Annonaceae	Leaves and stem bark	Ethanollic extract	----- --	In-vivo	LD90 (ppm): For adult= 6.21 For egg= 1	80 % mortality rate at 60ppm
Brazil	<i>Annona salmani</i>	Annonaceae	Leaves	Ethanollic extract	----- --	In-vivo	LD90 (ppm): For adult= 0.66 For egg= 1	100 % mortality rate
Brazil	<i>Annona squamosa</i>	Annonaceae	Seed, stem, root, and leaves	Ethanollic extract	----- --	In-vivo	LD90 (ppm): For adult= 8.55	100 % mortality rate

								For egg= 1		
Brazil	Euphorbia splendens	Euphorbiaceae	Latex	Aqueous solution	----- --	In-vivo	LD50= 0.60ppm LD90= 0.83 ppm	Extract proved toxic against snail sp.	[123]	
Iraq	Cucumis melo	Cucurbitaceae	Leaves and stem	Aquatic extract	----- --	In-vivo	LC50 (g/L): Leaves=40, Stem=50	15 % mortality rate with leaves extract while 12.5 % mortality rate with stem extract	[124]	
Iraq	Thymus Vulgaris	Lamiaceae	Leaves	Aqueous extract	----- --	In-vitro	LC50= 18.7 g/L	50 % mortality at 18.7g/L	[125]	
Iraq	Ricinus Communis	Euphorbiaceae	Leaves and bark	Aqueous extract	----- --	In-vivo	LC50 for 50 g/L Bark=191% Leaves= 32%	Abnormal behavioral patterns	[126]	
France	Croton campestris	Euphorbiaceae	Root and bark	dichloromethanolic extract	velamone, velamolone and velamolone acetate	In-vivo	LD100 =50 ppm	100 % mortality rate	[127]	
France	Caryopteris clandonensis	Lamiaceae	Root and bark	Chloroform extract	Fuerstione, α -Caryopterone and 15-	In-vivo	LD100= <3 ppm	100 % mortality rate	[128]	

						deoxyfur estione				
United Kingdom	Euphorbia myrsinites	Euphorbiaceae	Leaves and stem	Aqueous extract	----- --	In-vivo	LC50 = 8.9 ppm	75 % reduced activity and 100 % mortality rate	[129]	
Belgium	Maesa ceolata	Myrsinaceae	Leaves	Aqueous extract	Saponin	In-vivo	-----	>70 % mortality rate	[130]	
Germany	Jatropha curcas	Euphorbiaceae	Seed	Methanolic extract	----- --	In-vitro	LC100 =25 ppm	100 % mortality rate	[131]	
Egypt	Nigella sativa	Ranunculaceae	Seed	Oil	----- --	In-vitro	LC50= 5.8 ppm	Laceration of vacuolated tissues in foot region	[132]	
Egypt	Pelargonium graveolens	Geraniaceae	Seed	Oil	----- -	In-vitro	LC50= 15.0 ppm	Significant miracicidal and cercaricidal activity		
Egypt	Azadirachta indica	Meliaceae	Seed	Oil	----- -	In-vitro	LC50= 28.0 ppm	Formation of vacuoles in cytoplasm of digestive glands		
Egypt	Euphorbia pulcherima	Euphorbiaceae	Whole plant	Methanolic extract	----- --	In-vivo	LC50= 43 ppm	After 3 hour, mortality rate of cercariae is 50 % at 15 ppm, 100 % mortality rate of miracidia at 100 ppm, 28.4 % reduction in total protein and 40 % decrease in lactate dehydrogenase	[133]	

Egypt	<i>Atriplex nummularia</i>	Amaranthaceae	Whole plant	Methanolic extract	----- --	In-vivo	LC50= 32 ppm	After 3 hour, mortality rate of cercariae is 70 % at 15 ppm, 100 % mortality rate of miracidia at 100 ppm, 51 % reduction in total protein and 62.1 % decrease in lactate dehydrogenase	
Mali	<i>Glinus oppositifolius</i>	Aizoaceae	Whole plant	Crude extract	12-Glucosyl-GO1	In-vivo	LC50 of Butanol extract= 64.3 ppm, LC50 of Ethanolic extract= 86.2 ppm	100 % mortality rate of both snail species at 200 ppm	[68]
Egypt	<i>Sesbania sesban</i>	Leguminosae	Leaves	Methanolic extract	----- --	In-vivo, in-vitro	LC50= 15 ppm	Reduced cercarial production and 44 % mortality after 48h	[134]
Nigeria	<i>Talinum triangulare</i>	Talinaceae	Root	Ethanollic extracts	----- --	In-vivo	LC50= 251 ppm	Ovicidal activity at <300 ppm	[135]
Egypt	<i>Adenium obesum</i>	Apocynaceae	Leaves	Methanolic extract	----- --	In-vivo	LC25=26.3 %	Reduced protein content in tissues	[136]
Sudan	<i>Cymbopogon nervatus</i>	Poaceae	Leaves	Essential oils	----- --	In-vivo	LD50 = 237.33 ppm	100 % mortality rate at 400 ppm	[80]
Sudan	<i>Boswellia papyrifera</i>	Burseraceae	Leaves	Essential oils	----- --	In-vivo	LD50= 311.05 ppm	90 % mortality rate at 350 ppm	

	Morocco	<i>Withania somnifera</i>	Solanaceae	Leaves and fruit	Aqueous and methanolic extract	Flavonoids, terpenes, alkaloids, saponins, tannins	In-vivo	LC50 = 94 ppm	Proved toxic to snail specie	[137]
	Morocco	<i>Withania frutescens</i>	Solanaceae	Root, leaves and fruit	Aqueous extract	Terpenes, alkaloids, tannins and saponins	In-vivo	-----	Proved toxic to snail specie	
	Morocco	<i>Solanum elaeagnifolium</i>	Solanaceae	Fruit, Root	Aqueous extract	Alkaloids, tannins and saponins	In-vivo	LC50 = 9.5 ppm	Potential molluscicide against snail	
	Morocco	<i>Silene cucubalus</i>	Caryophyllaceae	Root	Aqueous and n-butanol extract	Saponins, tannins and terpenes	In-vivo	-----	Potential molluscicide against snail	
	Morocco	<i>Spergularia marginata</i>	Caryophyllaceae	Root	Aqueous extract	Saponins	In-vivo	-----	Proved toxic to snail specie	
	Morocco	<i>Saponaria vaccaria</i>	Caryophyllaceae	Root	Aqueous extract	Saponins	In-vivo	-----	Potential molluscicide against snail	
	Sudan	<i>Azadirachta indica</i>	Meliaceae	Leaves and seeds	Aqueous extract	----- --	In-vivo	-----	75 % decrease in snail species after 5 days	[84]
	Egypt	<i>Commiphora molmol</i>	Bruseraceae	Whole plant	Oil extract	----- --	In-vivo	LD50= 50 ppm	100 % mortality rate	[53]

								LD90= 95 ppm		
Morocco	<i>Citrus aurantium</i> var. valencia	Rutaceae	Fruit	Essential oil	α -pinene, sabinene, myrcene and linionene	In-vivo		LC50= 0.28 ppm	Toxic against the snail sp.	[138]
Morocco	<i>Origanum compactum</i>	Lamiaceae	Aerial parts	Essential oil	Thymol, α -terpineol, γ -terpinene and α -pinene	In-vivo		LC50= 0.44 ppm	Toxic against the snail sp.	
Mali	<i>Cussonia barteri</i>	Araliaceae	Roots	Methanolic and aqueous extract	----- ---	In-vitro		-----	Extract proved toxic against snail sp.	[85]
Mali	<i>Glinus oppositifolius</i>	Aizoaceae	Whole plant	Methanolic and aqueous extract	----- ---	In-vitro		-----	Extract proved toxic against snail sp.	
Mali	<i>Lannea velutina</i>	Anacardiaceae	Leaves	Methanolic and aqueous extract	----- ---	In-vitro		-----	Extract proved toxic against snail sp.	
Egypt	<i>Commiphora molmol</i>	Burseraceae	Whole plant	Oil and resin extract	----- ---	In-vivo		LC50= 3 ppm	100 % mortality rate	[56]
Morocco	<i>Ruta chalepensis</i> L.	Rutaceae	Aerial part	Aqueous and ethanolic extract	Phenol, coumarin, flavanoid, alkaloid, sterol and	In-vivo		LC50= 1.41 ppm LC90= 2.23 ppm	Strong molluscicidal activity	[139]

						terpenoids				
Morocco	<i>Zygophyllum gaetulum</i> Emb.	Zygophyllaceae	Aerial part	Aqueous and ethanolic extract	and	Flavonoid and saponin	In-vivo	LC50= 8.69 ppm LC90= 16.82 ppm	High mortality rate	
Morocco	<i>Citrus bigaradia</i> Duhamel	Rutaceae	Leaves	Aqueous and ethanolic extract	and	Terpenoid, phenol, coumarins, flavanoid, alkaloid, sterol and terpenoid	In-vivo	LC50= 2.74 ppm LC90= 4.43 ppm	Toxic for adult snails	
Morocco	<i>Capparis spinosa</i> L.	Capparidaceae	Aerial part	Aqueous and ethanolic extract	and	Coumarin, alkaloid and sterol	In-vivo	LC50= 70.58 ppm LC90= 122 ppm	Strong molluscicidal activity	
Morocco	<i>Chenopodium ambrosioides</i> L.	Chenopodiaceae	Aerial part	Aqueous and ethanolic extract	and	Flavonoid, saponin and terpenoid	In-vivo	LC50= 1.41 ppm LC90= 2.23 ppm	Strong molluscicidal activity	
Morocco	<i>Artemisia herba alba</i> Asso.	Asteraceae	Aerial part	Aqueous and ethanolic extract	and	Phenol, flavanoid, alkaloid,	In-vivo	LC50= 62.94 ppm	Strong molluscicidal activity	

						sterol and terpenoid		LC90= 92 ppm	
Morocco	<i>Origanum compactum</i>	Lamiaceae	Aerial part	Aqueous and ethanolic extract	Phenol, flavanoid, saponin and terpenoid	In-vivo	LC50= 8.22 ppm LC90= 1.99 ppm	Strong molluscicidal activity	
Morocco	<i>Melia azedarach</i> L.	Meliaceae	Leaves	Aqueous and ethanolic extract	Flavanioid, alkaloid and terpene	In-vivo	LC50= 3.53 ppm LC90= 4.20 ppm	High mortality rate	
Morocco	<i>Delphinium staphisagria</i> L.	Ranunculaceae	Flower	Aqueous and ethanolic extract	Alkaloid and saponin	In-vivo	LC50= 3.5 ppm LC90= 8.39 ppm	Toxic for adult snails	
Morocco	<i>Nigella sativa</i>	Ranunculaceae	Aerial part	Aqueous and ethanolic extract	Coumarins, alkaloid, terpenoid, saponin and sterol	In-vivo	LC50= 106.7 ppm LC90= 140 ppm	Toxic for adult snails	
Morocco	<i>Zizyphus vulgaris</i> Lamk.	Rhamnaceae	Leaves	Aqueous and ethanolic extract	Phenol, saponin and alkaloid	In-vivo	LC50= 4.12 ppm LC90= 8.41 ppm	High mortality rate	

	Morocco	<i>Chrysanthemum viscidhirtum</i>	Asteraceae	Aerial part	Aqueous and ethanolic extract	Flavonoid and terpenoid	In-vivo	LC50=4.43 ppm LC90=8.39 ppm	Strong molluscicidal activity	
	Morocco	<i>Citrullus colocynthis</i> L.	Cucurbitaceae	Fruit, Seed	Aqueous and ethanolic extract	Sterol and terpenoid	In-vivo	LC50= 4.35 ppm LC90= 8.39 ppm	Strong molluscicidal activity	
	Morocco	<i>Lavandula stoechas</i> L.	Lamiaceae	Aerial Part	Aqueous and ethanolic extract	Phenol, flavanoid and terpenoid	In-vivo	LC50= 106.7 ppm LC90= 140 ppm	High mortality rate	
	China	<i>Solanum xanthocarpum</i>	Solanaceae	Fruit	Ethanolic extract	----- --	In-vivo	LC50 (mg/l): =1.35	100 % mortality rate	[91]
	Germany	<i>Jatropha curcas</i>	Euphorbiaceae	Seed	Methanolic extract	----- --	In-vitro	LC100= 1 ppm	100 % mortality rate of snail specie	[131]
<i>Bulinus globosus</i>	Zimbabwe	<i>Cucurbita maxima</i>	Cucurbitaceae	Seed	Aqueous and crude ethanolic extracts	----- --	In-vivo	LC50 (mg/ml) for: Aqueous extracts on <i>B. globosus</i> adults= 0.004 Ethanolic extracts on	100 % mortality rate at 2 mg/mL	[59]

								B. globosus adults= 0.19		
	Nigeria	Allium cepa, Allium sativum	Amaryllidaceae	Bulb and cloves	Methanolic extract	----- ---	In-vivo	LC50 (60 mg/l): For Allium cepa=15; For Allium sativum= 19.37	Mortality is higher in case of Allium sativum than Allium sepa and highest mortality rate is observed in 42.15 mg/l	[140]
	Nigeria	Balanites aegyptiaca	Zygophyllaceae	Stem and bark	Aqueous solution	2-methoxy-4-vinylphenol, benzaldehyde, phenol, 2-methoxy-4-(2-propenyl)-acetate, ethanone, 1-(4hydroxyphenyl)-, alpha-d-6,3-furanose, 4-(3-hydroxy-1-	In-vivo	LD50 (mg/L): For juvenile= 5.7 For adult= 6.2 LD90 (mg/L): For juvenile= 6.8 For adult= 7.2	100 % mortality rate of both snail species at 40mg/L	[67]

						propenyl)-2-methoxy phenol, hexadecanoic acid, 2-butanone, 9-octadecanoic acid, octadecanoic acid, 13-octadecenal, tetradecanoic, 9-octadecanamide and di-n-octylphthalate				
	Nigeria	Rhizophora mangle	Rhizophoraceae	Leaves	Methanolic extract	Saponins, glycosides, tannins, alkaloids and flavanoids	In-vivo	LC50 = 87.50 ppm	90 % mortality rate at 200 ppm	[141]
	Nigeria	Rhizophora racemosa	Rhizophoraceae	Leaves	Methanolic extract	Saponins, glycosides, tannins, alkaloids	In-vivo	LC50 = 125 ppm	100 % mortality rate at 250 ppm	

						and flavanoids				
Nigeria	<i>Avicennia germinans</i>	Verbenaceae	Leaves	Methanolic extract	Saponins, glycosides, tannins, alkaloid and flavanoids	In-vivo	LC50= 87.50 ppm	80 % mortality rate at 150 ppm		
Nigeria	<i>Languncularia racemosa</i>	Combretaceae	Leaves	Methanolic extract	Saponins, glycosides, tannins, alkaloid and flavanoids	In-vivo	LC50 = 102.09 ppm	100 % mortality rate at 50 ppm		
Nigeria	<i>Azadirachta indica</i>	Meliaceae	Root and bark	Ethanol, chloroform, acetone and n-hexane extract	Triterpenes, flavanoids, glycosides and tannins	In-vivo	LC50 = 0.35 ppm	100 % mortality rate	[142]	
Nigeria	<i>Talinum triangulare</i>	Portulacaceae	Leaves and root	Ethanol extract	-----	In-vivo	LC50= 125.89 ppm	100 % mortality at 160 ppm (root) and 640 ppm (leaves)	[143]	
Nigeria	<i>Azadirachta indica</i>	Meliaceae	Leaves, seed and bark	Methanol and decoction extract	Saponin, tannin, flavanoid	In-vitro	-----	100 % mortality rate after 24 h	[144]	

						and glycoside				
	Nigeria	<i>Hyptis suaveolens</i>	Lamiaceae	Leaves	Ethanollic extracts	----- --	In-vivo	At egg stage LC50= 0.614 ppm; At adult stage LC50= 0.077 ppm	80 % mortality rate at 0.8 ppm	[145]
	Nigeria	<i>Securidaca longepedunculata</i> , <i>Tephrosia brateolata</i>	Polygalaceae, Leguminosae	Root , leaves, stem and bark	Ethanollic and methanollic extract	----- --	In-vivo	With ethanollic extract: LC50 (ppm) leaf= 0.15 bark = 0.19 Root= 0.18 With methanollic extract: LC50 (ppm) leaf = 0.55 bark= 0.60 root= 0.21	100 % mortality rate	[146]
	Nigeria	<i>Carica papaya</i>	Caricaceae	Leaves	Ethanollic extract	----- --	In-vivo	LC50 = 619.1 ppm LC90= 1180.7 ppm	100 % mortality rate	[79]

	Nigeria	Terminalia catappa	Combretaceae	Leaves	Ethanol extract	----- --	In-vivo	LC50 = 1095.7 ppm LC90 = 1874.9 ppm	100 % mortality rate	
Oncomelania hupensis	China	Camellia oleifera	Theaceae	Seed	Aqueous solution	Tea-seed distilled saponin	In-vivo	LD50 at 24 hr in laboratory = 0.701 mg/L	Mortality rate is 72-98 %	[147]
	China	Macleaya cordata	Papaveraceae	Fruit	Aqueous solution	Quaternary benzo[c]phenanthridine alkaloids (sanguinarine and chelerythrine) and protopine alkaloids (protopine and α -allocryptopine)	In-vivo, In-vitro	LC50=1.29 mg/L; LC90=2.92 mg/L	Decrease in glutathione transferase (GST), carboxylesterase, acid phosphatase and alkaline phosphatase activity	[148]
	Philippines	Jatropha curcas L.	Euphorbiaceae	Leaves	Petroleum ether extract	Alkaloids	In-vivo	LD50 = 0.45 mg/mL	80 % mortality at 1.5 mg/mL	[149]

						tannins, terpenoids, flavonoids and saponins				
China	<i>Macleaya cordata</i> (Wild) R.	Papaveraceae	Leaves	Aqueous solution	Alkaloids	In-vivo	LC50 = 6.35 mg/L LC90 = 121.23 mg/L	Mortality rate is 73 % at 25 mg/L, Increase in alanine aminotransferase, aspartate transaminase and decrease in esterase isozyme	[150]	
China	<i>Buddleja lindleyana</i>	Loganiaceae	Whole plant	Ethanollic extract	Acacetin-7-rutinoside	In-vivo	LC50 = 36:12 mg/L	100 % mortality at 70 mg/L	[151]	
China	<i>Cinnamomum camphora</i>	Lauraceae	Leaves	-----	Linalool, α -terpineol, 4-terpineol, camphora, β -phellandrene, 1,8-cineole and spathulenol	In-vitro	LC50 = 0.25 mg/L	Disfigurement noted in hepatopancrease and gills	[152]	

	China	Ginkgo biloba	Ginkgoaceae	Leaves and sarcotesta	Ethanollic extract	-----	In-vivo	LD50= 47.64 mg/L	Negatively affect the functioning of dihydronicotinamide adenine dinucleotide, cytochrome c oxidase, cytochrome b and (NADH) dehydrogenase, adenosine triphosphate (ATP) synthase	[153]
	China	Pulsatilla chinensis	Ranunculaceae	Whole plant	Alcohollic extract	Pulchinoside A3 and Pulchinoside B	In-vitro	LC50 =0.48 mg/L	Significant inhibition of cholinesterase, aspartate transaminase and alkaline phosphatase	[154]
	China	Sapium sebiferum	Euphorbiaceae	Leaves, branch, root and seed	Aqeous, ethyl acetate and ethnolic extract	----- ---	In-vivo	-----	Decreased protein content in liver and 100 % mortality rate	[155]
	China	Macleaya cordata	Papaveraceae	Fruit	Dechlorinized aqeous extract	Bisulfate s of sanguinarine, total alkaloids and chelerythrine	In-vivo	LC50= 0.19 mg/L	100 % mortality rate at 72h	[156]

China	<i>Eomecon chionantha</i>	Papaveraceae	Root stalk	Alcoholic extract	Alkaloids	In-vivo	-----	100 % mortality rate at 2.5 mg/l and increased amount of alanine aminotransferase	[157]
China	<i>Ginkgo biloba</i>	Ginkgoaceae	Leaves	Ethanollic and aqeous extract	----- --	In-vivo	-----	80 % mortality rate after 24 h, decreased total protein content and glycogen	[158]
China	<i>Torreya grandis</i>	Taxaceae	Leaves	Ethanollic and aqeous extract	----- --	In-vivo	-----	72 % mortality rate after 72 h	
China	<i>Ailanthus altissima</i>	Simaroubaceae	Leaves	Ethanollic and aqeous extract	----- --	In-vivo	-----	40 % mortality rate after 72 h	
China	<i>Peucedanum praerutorum</i>	Apiaceae	Root	Ethanollic and aqeous extract	----- --	In-vivo	-----	83 % mortality rate after 72 h	
China	<i>Herba agrimoniae</i>	Rosaceae	Whole plant	Ethanollic and aqeous extract	----- --	In-vivo	-----	80 % mortality rate after 72 h	
China	<i>Buddleja lindleyana</i>	Scrophulariaceae	Leaves	Ethanollic extract	----- --	In-vivo	LC50 = 39.91 mg/L LC90 = 59.28 mg/L	96 % mortality rate after 48 h	[159]
China	<i>Eupatorium adenophorum</i>	Asteraceae	Leaves, stem and root	Aqueous extracts	----- --	In-vivo	-----	100 % mortality rate after 58 h	[160]

	China	Ginkgo biloba	Ginkgoaceae	Seed	Petroleum ether extracts	Ginkgolic acids	In-vivo	LD50 = 7.81ppm	100 % mortality rate after 48h	[161]
	China	Arisaema erubescens	Araceae	Tubers	Ethanollic extract	----- --	In-vitro	LC50= 60.2 mg/L LC90= 145.6 mg/L	Increased activity of <i>alkaline phosphatase and choline esterase</i>	[162]
	China	Ginkgo biloba	Ginkgoaceae	Seed	Benzinum extracts	----- --	In-vivo	LC50= 0.65 mg/L LC90= 5.5 mg/L	Decreased activity of alkaline phosphatase, choline esterase, malic dehydrogenase and alanine aminotransferase	[163]
	China	Solanum xanthocarpum	Solanaceae	Fruit	Ethanollic extract	----- --	In-vivo	LC50 = 0.62 mg/l	100 % mortality rate	[164]
Bulinus rholfsi	Nigeria	Rhizophora mangle	Rhizophoraceae	Leaves	Methanollic extract	Saponins, glycosides, tannins, alkaloid and flavanoids	In-vivo	LC50 = 108.22 ppm	90 % mortality rate at 200 ppm	[141]
	Nigeria	Rhizophora racemosa	Rhizophoraceae	Leaves	Methanollic extract	Saponins, glycosides, tannins, alkaloid and	In-vivo	LC50= 83.51 ppm	100 % mortality rate at 250 ppm	

						flavonoid s				
	Nigeria	Avicennia germinans	Verbenaceae	Leaves	Methanolic extract	Saponins, glycoside s, tannins, alkaloid and flavonoid s	In-vivo	LC50 = 123.74 ppm	80 % mortality rate at 150 ppm	
	Nigeria	Languncularia racemosa	Combretaceae	Leaves	Methanolic extract	Saponins, glycoside s, tannins, alkaloid and flavonoid s	In-vivo	LC50= 152.03 ppm	100 % mortality rate at 50 ppm	
Bulinus natalensis	Germany	Jatropha curcas	Euphorbiaceae	Seed	Methanolic extract	----- ---	In-vitro	LC100= 1 ppm	100 % mortality rate of snail specie	[165]
Lymnaea luteola	India	Sapindus trifoliatus	Sapindaceae	fruit	Methanolic extract	----- ---	In-vitro	LC50= 50 ppm LC90= 87 ppm	80 % mortality rate	[166]
	India	Acacia concinna	Leguminosae	fruit	Methanolic extract	----- ---	In-vitro	LC50= 360 ppm LC90= 451 ppm	90 % mortality rate at 300 ppm	

	India	Madhuca indica	Sapotaceae	Seed	Methanolic extract	----- ---	In-vitro	LC50= 62 ppm LC90= 100 ppm	90 % mortality rate at 80 ppm	
	India	Phytolacca acinosa	Phytolaccaceae	fruit	Methanolic extract	----- ---	In-vitro	LC50= 193 ppm LC90= 399 ppm	80 % mortality rate	
Monacha cartusiana	Egypt	Syzygium aromaticum	Myrtaceae	Flower bud	Acetone and ethanol extract	----- -	In-vivo	LC50 =1.06; LC95=26.83	Excessive mucosal secretion, hyperactivity and 40-60 % mortality rate	[167]
	Egypt	Zingiber officinale	Zingiberaceae	Rhizome	Ethanol extract	----- ---	In-vitro, In-vivo	-----	90 % mortality at 28th day, Increased cell lysis and vacuole formation in digestive gland with disrupted calcium spherules and accumulation of pyknotic nuclei, increased levels of ALT, AST, α & β esterases and phenoloxidases, deformed egg and sperm	[168]
	Egypt	Balanites aegyptiaca	Zygophyllaceae	Fruit	Ethanol extract	Diosgenin and 3,5-spirostadiene	In-vivo	LC50= 0.256 ppm	73 % mortality rate	[169]

Monacha Obstructa	Egypt	<i>Ricinus communis</i>	Euphorbiaceae	Fruit and leaves	Aqueous and ethanolic extract	Rinisine, ricinoleic acid, linoleic acid and quinic acid	In-vivo	LC50= 0.215 LC90= 16.128	80 % mortality rate after 72 hr	[170]
	Egypt	<i>Anagallis arvensis</i> L.	Primulaceae	Leaves and fruit	Hexane extract	Malonic acid, aromadendrene, 2-tertbutyl-4-isopropyl-5-methylphenol and phosphoric acid tributyl ester	In-vivo	LC50 after 24h= 1.70 g/ml	Extract induced 34 % toxicity to adult snails	[171]
	Egypt	<i>Solanum nigrum</i> L.	Solanaceae	Leaves and fruit	Hexane extract	----- -	In-vivo	LC50 after 24h= 2.75 g/ml	Proved 75 % toxic	
	Egypt	<i>Cyperus longus</i> L.	Cyperaceae	Rhizome	Hexane extract	----- -	In-vivo	LC50 after 24h= 3.09 g/ml	Acted as bait (24.2 %) after 96 h	
	Saudi Arabia	<i>Adenium obesum</i>	Apocynaceae	Stem	Hydro-ethanolic extract	Cerberin and neriifolin	In-vivo	LD50 (µg/g):	Toxicity index of Cerberin is 79.81 while of neriifolin is 100	[172]

								Of Cerberin = 4.3 Of neriifolin = 5.39		
Indoplanorbis exustus	India	Artemisia annua	Asteraceae	Leaves	Crude extract	----- -	In-vitro	LC50= 5.9 ppm	Abnormal behavior (rapid movement)	[173]
	India	Leucas aspera, Parthenium hysterophorus	Lamiaceae, Asteraceae	Leaves	Aqueous solution	----- ----	In-vivo	-----	90 % mortality rate	[174]
Monacha cantiana	Saudi Arabia	Calotropis procera, Adenium arabicum	Apocynaceae	Latex and stem	Cardenolide and methonyl extract	Glycosides	In-vivo	LD50 values 12.62 mg/kg for cardenolide LD50 for methomyl was 116.62 mg/kg	80 % mortality rate	[175]
Lanistes ovum	Nigeria	Senna occidentalis	Leguminosae	Leaves	Ethanollic extract	Tannins, inulin, flavanoids and alkaloids	In-vitro	LC50=83.67 mg/L	100 % mortality rate after 96h	[176]
	Nigeria	Vernonia amygdalina	Asteraceae	Leaves	Ethanollic extract	Tannins, alkaloids, flavanoid	In-vitro	LC50=614.288 mg/L	100 % mortality rate after 96h	

						s and saponin				
	Nigeria	Khaya senegalensis	Meliaceae	Leaves	Ethanollic extract	Tannins, flavanoids, alkaloids, saponins and inulin	In-vitro	LC50=395.58 mg/L	Inhibition of alpha-glucosidase and reduced glucose level	
Physa acuta	Iraq	Melia azadirachta	Meliaceae	Fruit	Fruit extract	----- -	In-vivo	At 24th hr, LC50=10 ppm ; LC90 4.4 ppm	Reduced juvenile movement, Irregular heartbeat and reduced to 58 beats/min from 88 beats/min, 100 % mortality rate at 5 ppm	[177]
Cornu aspersum	Lebanon	Styrax officinalis	Styracaceae	Fruit	Ethanollic extract	Saponins, triterpenes and tannins	In-vivo	-----	Disintegrated membrane tissues	[178]
Subulina octona	Brazil	Mikania glomerata	Asteraceae	Leaves	Aqeous extract	Tannins	In-vivo	LC50=45 mg/mL	100% mortality rate at 60th day	[179]
	Brazil	Bidens pilosa Linné	Asteraceae	Aerial parts	Aqeous extract	Flavonoids and saponin	In-vivo	LC50=51.4 mg/mL; LC90=74.1 mg/mL	Reduced hatchability to 61 %, 100 % mortality rate after 48 h	[180]
Lanistes lybicus	Nigeria	Erythrophleum suaveolens	Leguminosae	Stem and bark	Ethanollic extract	Saponin	In-vivo	38.74 µg/ml	100 % mortality rate at 50 µg/ml	[181]
Physa marmorata	Brazil	Moringa oleifera	Moringaceae	Seed	Aqeous extract	----- -	In-vivo	P. marmorata LC50	Toxic for Physa marmorata	[107]

								=0.339 g/l; LC90 =0.789 g/l		
Bradybaena similaris	Brazil	Solanum paniculatum	Solanaceae	Leaves	Aqueous extract	----- -	In-vivo	-----	75 % mortality rate after 24 h, significant reduction of glucose, glycogen and galactogen	[182]
Bulinus sp.	Kenya	Balanites aegyptiaca	Balanitaceae	Leaves, root, stem and fruit	Methanolic extract	----- -	In-vitro	LD50= 2.0931	38 % mortality rate	[183]
	Kenya	Phytolacca dodecandra	Phytolaccaceae	Leaves, root, stem and fruit	Methanolic extract	----- -	In-vitro	LD50= 2.5332	32 % mortality rate	
	Kenya	Phytolacca octandra	Phytolaccaceae	Leaves, root, stem and fruit	Methanolic extract	----- -	In-vitro	LD50= 2.2627	36 % mortality rate	
	Nigeria	Moringa oleifera	Moringaceae	Seeds	Aqueous extract	----- -	In-vivo	-----	100 % mortality rate at 2000 ppm	[184]
Biomphalaria juvenile snails and Bulinus adult snails	Zimbabwe	Curcubita maxima	Cucurbitaceae	Seed	Aqueous extract	----- -	In-vivo	LC50 (mg/ml): On Biomphalaria juvenile= 0.002 On Bulinus adult = 0.004	100 % mortality rate	[185]

Different snail sp.	Sudan	Khaya grandifoliola	Meliaceae	Bark	Aqueous and ethanolic extract	----- -	In-vivo	-----	100 % mortality rate at 1 g/L	[186]
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A Most of the plants exhibiting molluscicidal activity belonged to the families Euphorbiaceae (9.31 %); Solanaceae (7.16 %); Asteraceae (6.09 %); Leguminosae (5.73 %); Lamiaceae (4.3 %); Meliaceae (3.94 %); Myrtaceae and Annonaceae (2.86 %);

Rutaceae and Zygophyllaceae (2.5 %); Moringaceae, Combretaceae, and Araliaceae (2.15 %), as presented in Figure 2. A number of other families ($n < 5$; < 1.79 %; Table 1) also included plants showing efficacy against various snail species.

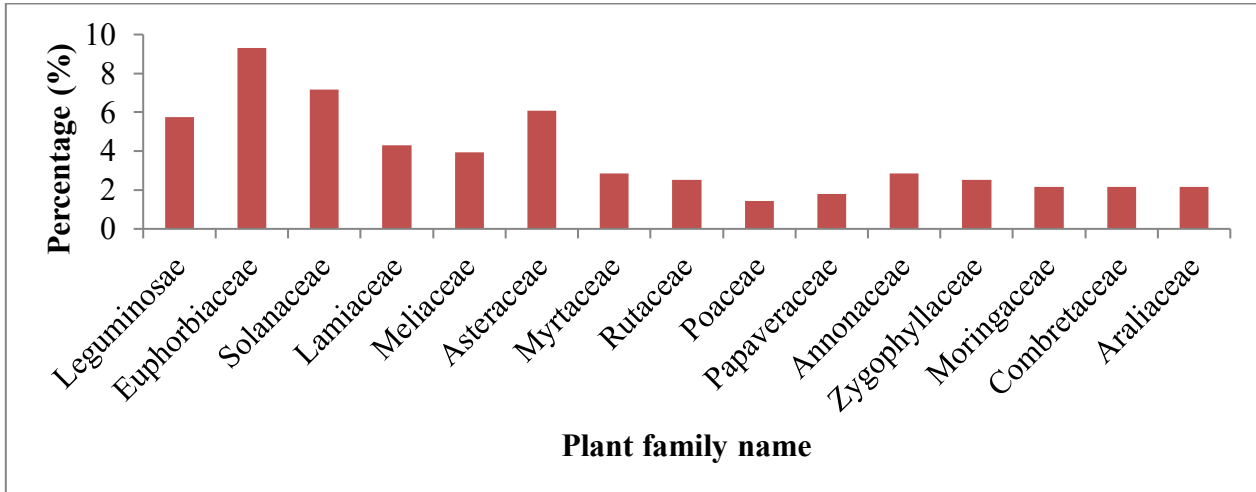


Figure 2: Major plant families exhibiting molluscicidal activity against intermediate snail hosts of schistosomes.

As shown in Figure 3, the key plant parts used to assess molluscicidal activity included leaves (48.74 %), fruit (15.05 %), root (14.69 %), stem (12.54 %), seed (12.9 %), bark (10 %), whole plant (9.31 %), and flower (2.15 %).

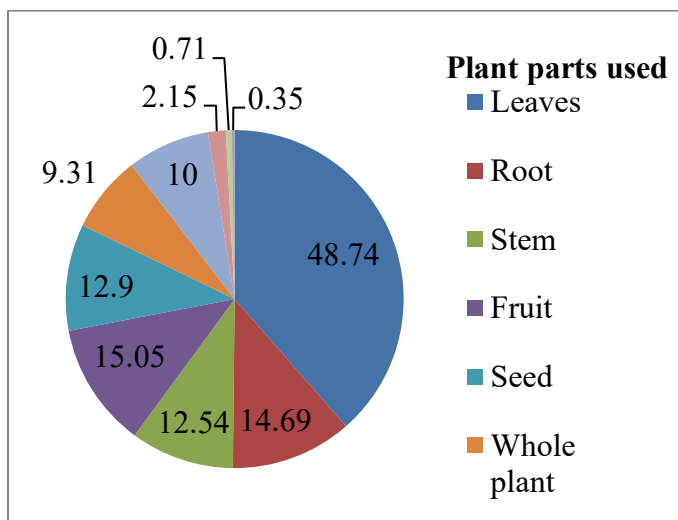


Figure 3: Pie chart depicting the percentage of plant parts used to assess molluscicidal activity.

Discussion

Plant-based molluscicides are receiving increasing recognition because of their higher biodegradability and low cost [63]. These molluscicides include plant secondary metabolites belonging to the category's tannins, glycosides, terpenoids, alkaloids, and phenols. Some of the important phytoconstituents are described below. Saponins are chemical compounds belonging to the category of glycosides. They are further divided into monodesmosidic and bidesmosidic saponins and are effective molluscicides because of their toxicity. One of the studies [18] indicated that the killing effect of *Phytolacca dodecandra* on snail species was attributed to saponins. In another study [187], saponins present in *Ocimum americanum* increased the mortality rate of *Biomphalaria pfeifferi*. Moreover, bidesmosidic saponins extracted from *Yucca desmettiana* leaves effectively reduced the occurrence of *Biomphalaria alexandrina*; the lethal concentration (LC₅₀) after 24 h of exposure was 68 mg/L [41].

The concept of essential oils (EOs) is increasingly gaining popularity since it has the potential to perform a wide range of beneficial functions such as combating bacteria, alleviating inflammation, and killing parasites, in particular. Recent research indicates that *Schistosoma* worms can be prevented to grow by using oils of various plants. Indicatively, A lipid extract of *Plectranthus neochilus* is very potent in worm-killing, which is primarily due to the presence of 2-caryophyllene and 2-pinene [188]. Likewise, oil of *Dysphania ambrosioides* is able to kill adult *Schistosoma mansoni* worms when placed in a laboratory [189]. This implies that we must continue to screen EOs to identify additional plant chemicals effective against

schistosomiasis. Similar effects have been found with other EOs, e.g. *Mentha x villosa* and *Foeniculum vulgare*, which is why it is believed that EOs may be natural remedies [190]. Analyzing EOs derived from plants can inform not only whether they are effective but also how they are: by rupturing the walls of cells of the parasite or by disrupting its most vital processes (Islam et al., 2020).

A study by [24], showed that alkaloids obtained from the leaf ethanolic extract of *Ziziphus spina-christi* led to a high apoptotic rate in *Biomphalaria alexandrina*. These alkaloids are organic compounds that are greatly soluble in chloroform and ethanol than water. They are present in every plant part and are proved highly toxic to the snails. Another study of Tripathi and Singh 2000, reported that isopelletierine isolated from *Punica granatum* significantly reduced the activity of *Lymnaea acuminata*. Alkaloids (sanguinarine) purified from *Macleaya cordata* also significantly decreased the activity of glutathione-S-transferase, carboxylesterase, and alkaline phosphatase in *Oncomelania hupensis* [150]. Flavonoids are flavone-based compounds in plants proven to exhibit molluscicidal activity. According to one of the studies [192], a well-known flavonoid (quercetin) isolated from the leaf ethanolic extract of *Polygonum senegalense* showed significant molluscicidal activity in *Biomphalaria pfeifferi* and *Biomphalaria glabrata* within 1 day of exposure. Flavonoids isolated from *Stryphnodendron adstringens* caused 90 % mortality in *Biomphalaria glabrata* [120]. Flavonoids are toxic for snail species, but flavone glycosides isolated from *Asparagus plumosus* were non-toxic [193].

Tannins are plant-based organic compounds divided into two groups: hydro-stable tannins and condensed tannins. Various plant species, including *Croton megalocarpus*, *Aloe*

secundiflora, *Moringa oleifera*, *Ipomea batatas*, *Chromolaena odorata*, *Ocimum americanum*, *Stryphnodendron adstringens* [107,117,187,194] are reported to contain tannins that are highly toxic to snails. Tannins present in *Persea Americana* enhanced the mortality rate of *Biomphalaria glabrata* embryos and caused lethargy in *Biomphalaria glabrata* adults [97]. Several plant species have been demonstrated to be effective against various snail species, but there are no sufficient data to determine their mechanism of action [195]. In 1998, WHO presented a report that emphasized the need to investigate the mode of action of molluscicidal plants, as they are economical, sustainable to use, and commonly available [196].

Plant molluscicides are classified on the basis of their mode of action into the following categories: enzyme inhibitors, neurotransmitter inhibitors (neurotoxins), stomach poisons, respiratory poisons, contact poisons, and growth inhibitors. The findings of some studies that investigated the mechanism of action of molluscicidal plants are discussed below: In recent years, a few studies have investigated the system of mollusks affected by molluscicides. [92] demonstrated that the latex of *Euphorbia cauducifolia* disrupted water absorption in *Biomphalaria glabrata*. The snail species that act as intermediate hosts of schistosomes are mainly in fresh water; therefore, the hemolymph of these snails is hyper-osmotic in nature. Consequently, it is crucial that the nutrients and water are in continuous equilibrium; their imbalance leads to reduced functioning and altered behavior (rapid and disoriented movement). It also causes tissue swelling, which makes snails more susceptible to plant molluscicides [8].

Plant molluscicides cause significant hormonal changes in snails, leading to a high mortality rate [14]. Enzymes such as acetylcholinesterase, succinate dehydrogenase, and alkaline phosphatase are used as indicators of plant molluscicidal activity. *Biomphalaria alexandrina* exposed to dry powdered *Datura stramonium* exhibited an increase in the levels of alanine transaminase and alkaline phosphatases [172]. Another study [49] showed that the methanolic extract of *Guayacum officinalis* decreased the levels of acetylcholinesterase and succinate dehydrogenase in *Biomphalaria alexandrina*. An aqueous extract of *Arisaema erubescens* was also shown to decrease the levels of superoxide dismutase and peroxidase in the liver of *Oncomelania hupensis*; in contrast, the malondialdehyde content increased, which indicates the oxidative stress damage caused by the plant extract, leading to snail mortality [162]. Molluscicides may also act by negatively affecting the growth and development of snails. The findings of Ibrahim et al. [20] indicated that cerium oxide nanocomposites isolated from *Moringa oleifera* significantly reduced the reproductive rate of *Biomphalaria alexandrina*, with an LC50 of 314.5 mg/L. The results of this study also indicated the deformation of hermaphrodite and digestive glands in the *Biomphalaria*

alexandrina. Certain flavonoids are highly toxic to schistosomes and others are not. This disparity is based on their structure, which influences the interaction with cell and their availability within the body. The hydroxyl groups of flavor compounds tend to make the flavonoids better antioxidants and antiparasites when they are present compared to when they are absent [197]. Hence, the research on the structure-activity relationship (SAR) of these compounds should be examined by scientists to inform future studies on flavonoid drugs against schistosomiasis.

Schistosomiasis also depends on the temperature of the surrounding environment to be spread. Research demonstrates that increased temperatures accelerate *Bulinus* snail life cycle and lead to more cercariae being released, and other research illustrates that high temperatures cause stress in the snails and lower their parasite-carrying capacity [198]. Such mixed evidence implies we require more ecological models to anticipate how schistosomiasis will evolve with different temperatures, particularly with the rising heat extremes with climate change.

When snails encounter a toxic substance, they produce more mucus in an attempt to excrete the noxious substance and detoxify their body [196]. The mucus forms a protective barrier around the skin and reduces the harmful effect of molluscicides [199]. Similarly, several other factors have been reported to decrease the molluscicidal activity of plants. The surface-to-volume ratio of snails is another factor responsible for the variation in the efficacy of plant molluscicides. According to [59], the effect of *Cucurbita maxima* extracts on *Biomphalaria pfeifferi* juveniles and adults are different. The LC50 values for adults and juveniles were 0.002 and 0.004 mg/mL, respectively. The reason is the less absorption of extract leads to more adult snail susceptibility towards toxic effect of molluscicides. Apart from these biological factors, certain chemical factors influence the susceptibility of snails to these plant extracts. For example, enzymatic analysis suggests that animals—more specifically, invertebrates—have mixed-function oxidases in their hepatopancreas that oxidize toxic chemical compounds to make them non-toxic [199].

Environmental factors such as light, temperature and extraction technique also affect the efficacy of plant-based molluscicides. A study by [112] indicated that an increase in temperature decreased the molluscicidal activity of *Euphorbia milli*. In contrast, an increase in temperature during phytochemical extraction helps these chemicals to easily penetrate the snail tissue [200]. Notably, [195] reported that the mortality rate of *Lymnea acuminata* treated with oleoresin decreased during winter.

Conclusion

This review highlights the significant potential of plant-derived compounds as eco-friendly molluscicides for controlling schistosomiasis. The evidence indicates that several plants and their phytochemicals exhibit strong molluscicidal activity, providing a promising alternative to synthetic chemicals. However, most studies remain limited to laboratory settings, and translation into large-scale field application is still scarce. Future research should focus on standardizing extraction methods, evaluating long-term safety and environmental impacts, and conducting field trials to validate laboratory findings. Collaborative efforts between researchers, policymakers, and public health authorities are essential to integrate plant-based molluscicides into schistosomiasis control programs. Such an approach could help reduce transmission while promoting sustainable and cost-effective disease management strategies.

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Conflicts of Interest

The authors declare no conflicts of interest.

References

- Nelwan ML. Schistosomiasis: Life Cycle, Diagnosis, and Control. *Curr Ther Res Clin Exp* 2019;91:5–9. <https://doi.org/10.1016/j.curtheres.2019.06.001>.
- Colley DG, Bustinduy AL, Secor WE, King CH. Human schistosomiasis. *The Lancet* 2014;383:2253–64. [https://doi.org/10.1016/S0140-6736\(13\)61949-2](https://doi.org/10.1016/S0140-6736(13)61949-2).
- Qi YX, Huang MR, Sun HY, Wu XY, Liu ZT, Lu DB. Prevalence of depressive symptoms in patients with advanced schistosomiasis in China: A systematic review and meta-analysis. *JournalsPlosOrg* 2024;18. <https://doi.org/10.1371/JOURNAL.PNTD.0012003>.
- Muhsin MA, Wang X, Kabole F, Zilabumba J, Yang K. The Indispensability of Snail Control for Accelerating Schistosomiasis Elimination: Evidence From Zanzibar. *Trop Med Infect Dis* 2022. <https://doi.org/10.3390/tropicalmed7110347>.
- M. Trienekens SC, Faust CL, Besigye F, Pickering L, Tukahebwa EM, Seeley J, et al. Variation in Water Contact Behaviour and Risk of *Schistosoma Mansoni* (Re)infection Among Ugandan School-Aged Children in an Area With Persistent High Endemicity. *Parasit Vectors* 2022. <https://doi.org/10.1186/s13071-021-05121-6>.
- Gong Y, Feng J, Luo Z-W, Xue J-B, Guo Z-Y, Zhang L, et al. Spatio-Temporal Heterogeneity of Schistosomiasis in China Based on Multi-Stage, Continuous Downscaling of Sentinel Monitoring 2021. <https://doi.org/10.21203/rs.3.rs-776529/v1>.
- Barbosa FS, Pereira da Costa D, Arruda F. New field observations on the competitive displacement between two species of planorbid snails inhabiting northeastern Brazil. *Mem Inst Oswaldo Cruz* 1981;76:361–6. <https://doi.org/10.1590/S0074-02761981000400004>.
- Allan F, Ame SM, Tian-Bi YNT, Hofkin B V., Webster BL, Diakité NR, et al. Snail-related contributions from the schistosomiasis consortium for operational research and evaluation program including xenomonitoring, focal mollusciciding, biological control, and modeling. *American Journal of Tropical Medicine and Hygiene* 2020;103:66–79. <https://doi.org/10.4269/ajtmh.19-0831>.
- Huot C, Clerissi C, Gourbal B, Galinier R, Duval D, Toulza E. Schistosomiasis Vector Snails and Their Microbiota Display a Phyllosymbiosis Pattern. *Front Microbiol* 2020;10:1–10. <https://doi.org/10.3389/fmicb.2019.03092>.
- Research MN-CT, 2019 undefined. Schistosomiasis: life cycle, diagnosis, and control. Elsevier 2019;91:5–9. <https://doi.org/10.1016/J.CURTHERES.2019.06.001>.
- Augusto R de C, de Mello-Silva CCC. Phytochemical molluscicides and schistosomiasis: What we know and what we still need to learn. *Vet Sci* 2018;5:1–9. <https://doi.org/10.3390/vetsci5040094>.
- World Health Organization. Progress report 2001–2011 and strategic plan 2012–2020. 2010.
- Inobaya M, Olveda R, Chau T, Olveda D, Ross A. Prevention and control of schistosomiasis: a current perspective. *Res Rep Trop Med* 2014;5:65–75. <https://doi.org/10.2147/rrtm.s44274>.
- Zheng L, Deng L, Zhong Y, Wang Y, Guo W, Fan X. Molluscicides against the snail-intermediate host of *Schistosoma*: a review. *Parasitol Res* 2021;120:3355–93. <https://doi.org/10.1007/s00436-021-07288-4>.
- Castle GD, Mills GA, Gravell A, Jones L, Townsend I, Cameron DG, et al. Review of the molluscicide metaldehyde in the environment. *Environ Sci (Camb)* 2017;3:415–28. <https://doi.org/10.1039/c7ew00039a>.
- He P, Wang W, Sanogo B, Zeng X, Sun X, Lv Z, et al. Molluscicidal activity and mechanism of toxicity of a novel salicylanilide ester derivative against *Biomphalaria* species. *Parasit Vectors* 2017;10:1–11. <https://doi.org/10.1186/s13071-017-2313-3>.
- Kashyap S, Khagta S, Guleria K, Arya V. Plants as Molluscicides : A recent update. *Int J Bot Stud* 2019;4:50–6.

18. Lemma A. Laboratory and field evaluation of the molluscicidal properties of *Phytolacca dodecandra*. Bull World Health Organ 1970;42:597–612.
19. Kloos H, McCullough FS. Plant Molluscicides. J Med Plant Res 1982;46:195–209.
20. Ibrahim AM, Mohamed F, Al-Quraishy S, Abdel-Baki AAS, Abdel-Tawab H. Green synthesis of Cerium oxide / *Moringa oleifera* seed extract nano-composite and its molluscicidal activities against *Biomphalaria alexandrina*. J King Saud Univ Sci 2021;33:101368. <https://doi.org/10.1016/j.jksus.2021.101368>.
21. Ibrahim AM, Saleh HA, Zayed KM. *Colchicum Ritchii* flower: a new molluscicidal plant for *Biomphalaria alexandrina* snails and the infective stages of *Schistosoma mansoni*. Molluscan Res 2021;41:289–97. <https://doi.org/10.1080/13235818.2021.2003982>.
22. Ibrahim AM, Sayed SSM. Assessment of the molluscicidal activity of the methanolic seed extracts of *Ziziphus spinachristi* and *Carica papaya* on immunological and molecular aspects of *Biomphalaria alexandrina* snails. Aquac Res 2021;52:2014–24. <https://doi.org/10.1111/are.15050>.
23. Abou El-Nour MF. Evaluation of molluscicidal, miracidicidal and cercaricidal activities of crude aqueous extracts of *origanum majorana*, *ziziphus spina-christi* and *salvia fruticosa* on *Schistosoma mansoni* and *Schistosoma haematobium*. Egypt J Aquat Biol Fish 2021;25:913–33. <https://doi.org/10.21608/ejabf.2021.173661>.
24. Ibrahim AM, Ghoname SI, Mansour SM, El-Dafrawy SM. Effect of some medicinal plant extracts as molluscicidal and apoptotic agents on *Biomphalaria Alexandrina* snails. Egypt J Aquat Biol Fish 2020;24:291–300. <https://doi.org/10.21608/ejabf.2020.80284>.
25. Rabia Ibrahim AB, Gouida MSO, Elsayed H, Attiyah S. Efficiency of Three Extracts of *Carica papaya* as Molluscicidal and Anti-schistosomal Agents against *Biomphalaria alexandrina* and *Schistosoma mansoni* by Flow Cytometry Efficiency of Three Extracts of *Carica papaya* as Molluscicidal and Anti-schistosomal Ag. J Pharm Res Int 2020;32:31–41. <https://doi.org/10.9734/JPRI/2020/v32i1130577>.
26. A. Saad AH, Azzam AM, Mostafa BB, El-said KM, Hanafy RA. Improvement molluscicidal activity of *Anagallis arvensis* extracted by copper oxide nanoparticles against *Biomphalaria alexandrina* snails Abdel. Egypt J Aquat Biol Fish 2019;23:27–41.
27. El-Khayat HMM, Metwally KM, Abououf NA. Histopathological and Ultrastructural Studies on *Biomphalaria Alexandrina* Snails Infected with *Schistosoma Mansoni* Miracidia and Treated with Plant Extracts. Egypt J Hosp Med 2018;71:2792–804. <https://doi.org/10.12816/0045846>.
28. Ibrahim AM, Ghoname SI. Experimental Parasitology Molluscicidal impacts of *Anagallis arvensis* aqueous extract on biological, hormonal, histological and molecular aspects of *Biomphalaria alexandrina* snails. Exp Parasitol 2018;192:36–41. <https://doi.org/10.1016/j.exppara.2018.07.014>.
29. Bahgat DM, Mossalem HS, Al-sayed E, Eldahshan OA, Singab ANB, Abu HM, et al. Influence of saponin fraction from *Albizia anthelmintica* on *Biomphalaria alexandrina* snail; the intermediate host of *Schistosoma mansoni* in Egypt. Egypt J Aquat Biol Fish 2018;22:231–40.
30. Mohamed AH, Osman GY, El-Eman MA, Abdel-Hamid H, E.M. Ali R. Deteriorations in biological and biochemical aspects of *Biomphalaria alexandrina* snails exposed to methanol extract of *Callistemon citrinus* leaves. J Egypt Soc Parasitol 2018;48:343–55.
31. El-Emam M, GY O, Abdel-Hamid H, Mohamed AH, R E M A. Determination of egg laying capacity, sex hormones and mortality of *Biomphalaria alexandrina* snails exposed to methanol extract from curcumin and the plants *Callistemon citrinus* and *Zingiber officinale*. J Biosci Appl Res 2017;3:97–109.
32. El-deeb FAA, Marie MS, Hasheesh WS, Sayed SSM. Factors affecting the molluscicidal activity of *Asparagus densiflorus* and *Oreopanax guatemalensis* plants and *Difenoconazole* fungicide on *Biomphalaria alexandrina* snails. Comp Clin Path 2016:1–9. <https://doi.org/10.1007/s00580-016-2263-8>.
33. Ghaly NS, Mina SA, Younis N. Schistosomicidal and molluscicidal activities of two Junipers species cultivated in Egypt and the chemical composition of their essential oils. Journal of Medicinal Plants Research 2016;10:47–53. <https://doi.org/10.5897/jmpr2015.5993>.
34. Hamed M, El-Amin S, Abdel-Fattah AS. Molluscicidal activity of some constituents isolated from *Cestrum purpureum*. Pharmacology Online 2015;2:59–72.
35. Ibrahim AM, Abdel-Gawad MM, El-Nahas HA, Osman NS. Studies on the Molluscicidal Activity of *Agave Angustifolia* and *Pittosporum tobira* on *Schistosomiasis* transmitting snails. Journal of Egyptian Society of Parasitology 2015;45:133–41.
36. El-sayed MM, El-nahas HA, El-Wakil EA. Molluscicidal Properties and Chemical Constituents of *Euphorbia peploides*. Australian Journal of Basic and Applied Sciences 2014;8:541–8.
37. Gohary E, Laila RA, Marwa GAM. Biochemical Effect of Three Molluscicide Baits Against the Two Land Snails, *Monacha cantiana* and *Eobania vermiculata* (Gastropoda: Helicidae). International Journal of Agricultural Research 2011;6:682–90.
38. Al-Sayed E, Hamid HA, Abu El Einin HM. Molluscicidal and antischistosomal activities of methanol extracts and isolated compounds from *Eucalyptus globulus* and *Melaleuca stypelioides*. Pharm Biol 2014;52:698–705. <https://doi.org/10.3109/13880209.2013.865240>.
39. Sheir SK, Mohamed AH, Osman GY, El-nabi SEH, Allam A. Delta Journal of Science Efficiency of Citrus limon extract on biological and molecular activities of *Biomphalaria alexandrina* snails. Delta Journal of Science 2013;36:186–200.
40. Abdel-haleem AA. Comparative karyological studies on the two Egyptian schistosome vectors, *Biomphalaria glabrata* and *B. alexandrina*, with reference to chromosomal aberrations due to *Za'ater* plant. International Journal of Academic and Scientific Research 2013;1:1–7.
41. Diab Y, Ioannou E, Emam A, Vagias C, Roussis V. Desmettianosides A and B, bisdesmosidic furostanol saponins with molluscicidal activity from *Yucca*

- desmettiana. Steroids 2012;77:686–90. <https://doi.org/10.1016/j.steroids.2012.02.014>.
42. Rizk MZ, Metwally NS, Hamed MA, Mohamed AM. Experimental Parasitology Correlation between steroid sex hormones , egg laying capacity and cercarial shedding in *Biomphalaria alexandrina* snails after treatment with *Haplophyllum tuberculatum*. Exp Parasitol 2012;132:171–9. <https://doi.org/10.1016/j.exppara.2012.06.011>.
 43. Mahmoud MB, Ibrahim WL, Abou-el-nour BM, El-emam MA, Youssef AA. Biological and biochemical parameters of *Biomphalaria alexandrina* snails exposed to the plants *Datura stramonium* and *Sesbania sesban* as water suspensions of their dry powder. Pestic Biochem Physiol 2011;99:96–104. <https://doi.org/10.1016/j.pestbp.2010.11.005>.
 44. Bakry FA, Mohamed RT, El-homossany K. Biological and biochemical responses of *Biomphalaria alexandrina* to some extracts of the plants *Solanum siniacum* and *Artemisia judaica* L . Pestic Biochem Physiol 2011;99:174–80. <https://doi.org/10.1016/j.pestbp.2010.12.001>.
 45. El-din AS, El-sayed K, Mahmoud M. Effect of ethanolic extract of *Dalbergia sissoo* plant parts on *Biomphalaria alexandrina* snail , the intermediate host of *Schistosoma mansoni*. Journal of Evolutionary Biology Research Vol 2011;3:95–100. <https://doi.org/10.5897/JEBR11.009>.
 46. Osman GY, Mohamed AM, Kader AA, Mohamed AA, Kader AA, Mohamed AA. Biological studies on *Biomphalaria alexandrina* snails treated with *Furcraea selloa marginata* plant (family : Agavaceae) and *Bacillus thuringiensis kurstaki* (Dipel-2x). J Appl Pharm Sci 2011;1:47–55.
 47. Bakry FA, Mohamed RT. Impact of *Euphorbia milii* latex on infectivity of *Schistosoma mansoni* larval stages to their hosts. Journal of Evolutionary Biology Research 2011;3:101–7. <https://doi.org/10.5897/JEBR11.015>.
 48. Hassan S, Abdel Rahman E, Abd El Monem AR. Molluscicidal activity of butanol fraction of *Meryta denhamii* flowers against *Lymnaea natalensis* and *Biomphalaria alexandrina*. Glob Vet 2010;4:15–21.
 49. Bakry FA. Use of some plant extracts to control *Biomphalaria alexandrina* snails with emphasis on some biological effects. Pestic Biochem Physiol 2009;95:159–65. <https://doi.org/10.1016/j.pestbp.2009.08.007>.
 50. Attia WY, El-Bolkiny YE, Al-Sharkawi IM, Mohamed SH. Efficacy of Mandarin (*Citrus Reticulata*) Peel Extract in the Control of *Schistosoma Mansoni* Larval Stage and Their Intermediate Hosts. The Egyptian Society of Experimental Biology 2009;5:247–54.
 51. Abdelgaleil SAM, El-Aswad AF, Nakatani M. Molluscicidal and anti-feedant activities of diterpenes from *Euphorbia paralias* L. Pest Manag Sci 2002;58:479–82. <https://doi.org/10.1002/ps.487>.
 52. Motawea S, El-Gilany A-H, Salama O. Preliminary study of the molluscicidal action of Myrrh. Journal of Environmental Science 2001;21:153–62.
 53. Allam AF, el-Sayad MH, Khalil SS. Laboratory assessment of the molluscicidal activity of *Commiphora molmol* (Myrrh) on *Biomphalaria alexandrina*, *Bulinus truncatus* and *Lymnaea cailliaudi*. J Egypt Soc Parasitol 2001;31:683–90.
 54. El-Sayed MM, El-Nahas HA. Molluscicidal Steroidal Saponins From *Yucca Elephantipes*. Zagazig Journal of Pharmacological Science 2001;10:41–8.
 55. Allam AF. Evaluation of different means of control of snail intermediate host of *Schistosoma mansoni*. J Egypt Soc Parasitol 2000;30:441–50.
 56. Masoud AM, Fawzy SM, Salama OM. Laboratory studies on the molluscicidal and cercaricidal activities of *Commiphora molmol*. Egypt J Aquat Biol Fish 2000;4:251–66.
 57. El-Ghaban AG, Eldiasty JG, M. H. M. Kamel O. Evaluation of *Rosmarinus* species extracted by different solvents against mosquito larvae, *Biomphalaria* species and different pathogenic bacteria. J Adv Sci Res 2015;6:44–9.
 58. Jangwan JS, Aquino RP, Mencherini T, Picerno P, Singh R. Chemical constituents of ethanol extract of leaves and molluscicidal activity of crude extracts from *Vitex trifolia* Linn . Nerva Polonica 2013;59:19–32. <https://doi.org/10.2478/hepo-2013-0021>.
 59. Mtemeli FL, Walter I, Tinago T, Shoko R. An assessment of the molluscicidal potential of *Cucurbita maxima* seed extracts on *Biomphalaria pfeifferi* and *Bulinus globosus* snails. All Life 2021;14:244–55. <https://doi.org/10.1080/26895293.2021.1901788>.
 60. Isa M. Laboratory Assessment of Molluscicidal and Cercaricidal Activities of *Balanites aegyptiaca* against Vectors of Schistosomiasis (*Biomphalaria pfeifferi*). International Journal of Research and Review 2021;8:479–86. <https://doi.org/10.52403/ijrr.20210558>.
 61. Albagouri AH, Elegami AA, Koko WS. Molluscicidal Activity of Some Sudanese Medicinal Plants Against Vector Snails of *Schistosoma mansoni*. Nile Journal for Agricultural Sciences 2021;6:36–47.
 62. Abdelmageed E, Bushara HO, Abdelgadir M. Evidence of biological activity of *Pulicaria crispa* on *Biomphalaria pfeifferi* host snails of *Schistosoma mansoni*. Biosciences Biotechnology Research Communications 2017;11:55–9. <https://doi.org/10.21786/bbrc/11.1/8>.
 63. Mandefro B, Mereta ST, Tariku Y, Ambelu A. Molluscicidal effect of *Achyranthes aspera* L. (Amaranthaceae) aqueous extract on adult snails of *Biomphalaria pfeifferi* and *Lymnaea natalensis*. Infect Dis Poverty 2017;6:1–5. <https://doi.org/10.1186/s40249-017-0349-4>.
 64. Elsareh F, Abdalla R, Abdalla E. The effect of aqueous leaves extract of *Solenostemma argel* (Del Hayne) on egg masses and neonates of *Biomphalaria pfeifferi* snails. Journal of Medicinal Plants 2016;4:271–4.
 65. Mwonga KB, Waniki NE, Dorcas YS, Piero NM. Molluscicidal Effects of Aqueous Extracts of Selected Medicinal Plants from Makueni County, Kenya. Pharm Anal Acta 2015;6:1–4. <https://doi.org/10.4172/2153-2435.1000445>.
 66. Ahmed EA, Babiker OF, Abdalla RM. Molluscicidal Activity of Aqueous Leave Extract of *Solenostemma argel* (Del Hayne) on *Biomphalaria pfeifferi* Snails I : INTRODUCTION. Journal of Basic and Applied Scientific Research 2014;4:179–84.
 67. Hena JS, Sindama A, Kulawe D. Gas Chromatography Mass Spectrometric Analysis and Molluscicidal Effect of Crude Saponin from *Balanites aegyptiaca* (L .) Del . J Adv

- Biol Biotechnol 2017;15:1–7. <https://doi.org/10.9734/JABB/2017/28802>.
68. Denou A, Togola A, Inngierdingen KT, Diallo D, Paulsen BS. Laboratory Assessment of Molluscicidal Activity of *Glinus Oppositifolius* (L .) Aug . DC . Related papers Laboratory Assessment of Molluscicidal Activity of. International Journal of New Technology and Research 2016;2:123–6.
 69. Kiros G, Erko B, Giday M, Mekonnen Y. Laboratory assessment of molluscicidal and cercariacidal effects of *Glinus lotoides* fruits. BMC Res Notes 2014;7:1–7. <https://doi.org/10.1186/1756-0500-7-220>.
 70. Molla E, Giday M, Erko B. Laboratory assessment of the molluscicidal and cercariacidal activities of *Balanites aegyptiaca*. Asian Pac J Trop Biomed 2013;3:657–62. [https://doi.org/10.1016/S2221-1691\(13\)60132-X](https://doi.org/10.1016/S2221-1691(13)60132-X).
 71. Michael ES, Yole D, Musila FM, Kutima H, Kareru P. Assessment of Molluscicidal, Cercericial and Miracidal Activities of Crude Extracts of *Azadirachta indica* and *Entada leptostachya*. J Biol Agric Healthc 2013;3:11–8.
 72. Otarigho B, Morenikeji OA. Efficacy of aqueous and ethanolic extracts of leaves of *Chromolaena odorata* as molluscicide against different developmental stages of *Biomphalaria pfeifferi*. Afr J Biotechnol 2013;12:438–44. <https://doi.org/10.5897/AJB12.2156>.
 73. Oledibe PA, Morenikeji OA, Benson O. Zoology and Ecology Molluscicidal potency of *Ficus exasperata* (Vahl) against juvenile and adult *Biomphalaria pfeifferi*. Zoology and Ecology 2013;23:147–56. <https://doi.org/10.1080/21658005.2013.797150>.
 74. Benson O, Morenikeji OA. Molluscicidal Effects of Aqueous and Ethanolic Extracts of Lemongrass (*Cymbopogon Citratus*) Leaf against the Different Developmental Stages of *Biomphalaria Pfeifferi*. New York Science Journal 2012;5:70–7.
 75. Benson O. Efficacy of *Chromolaena odorata* leaf extracts on *Biomphalaria pfeifferi* eggs in control of Schistosomiasis. Zoology and Ecology 2012;22:236–9. <https://doi.org/10.1080/21658005.2012.745269>.
 76. Ismail MA, Koko WS, Osman EE, Dahab MM, Garbi MI, Alsadeg AM, et al. Molluscicidal Activity of *Acacia seyal* (Dell) Bark Methanolic Extract Against *Biomphalaria pfeifferi* Snails. International Biological and Biomedical Sciences Journal 2016;2:73–9.
 77. Ukwandu N, Bdaibo A, Okorie T, Nmorsi O. Molluscicidal effect of *Piper guineense*. African Journal of Complementary Alternative Medicine 2011;8:447–51. <https://doi.org/10.4314/ajtcam.v8i4.17>.
 78. Adenusi AA, Odaibo AB. A laboratory assessment of the potential molluscicidal activity of some Nigerian plant species used as anthelmintics. Afr J Aquat Sci 2010;35:251–8. <https://doi.org/10.2989/16085914.2010.545506>.
 79. Adentuji V o., Salawu OT. Efficacy of ethanolic leaf extracts of *Carica papaya* and *Terminalia catappa* as molluscicides against the snail intermediate hosts of schistosomiasis. Journal of Medicinal Plants 2010;4:2348–52. <https://doi.org/10.5897/JMPR10.468>.
 80. El-kamali HH, El-Nour OR, Khalid SA. Molluscicidal Activity of the Essential Oils of *Cymbopogon nervatus* Leaves and *Boswellia papyrifera* Resins. Current Research Journal of Biological Sciences 2010;2:139–42.
 81. Adenusi AA, Odaibo AB. Effects of varying concentrations of the crude aqueous and ethanolic extracts of *Dalbergia Sisso* plant parts on *Biomphalaria pfeifferi* egg masses. African Journal of Traditional, Complementary and Alternative Medicines 2009;6:139–49.
 82. Adenusi AA, Odaibo AB. Laboratory assessment of molluscicidal activity of crude aqueous and ethanolic extracts of *Dalbergia sissoo* plant parts against *Biomphalaria pfeifferi*. Travel Med Infect Dis 2008;6:219–27. <https://doi.org/10.1016/j.tmaid.2007.12.004>.
 83. Ojewole JAO, Nundkumar N, Adewunmi CO. MOLLUSCICIDAL, CERCARIACIDAL, LARVICIDAL AND ANTIPLASMODIAL PROPERTIES OF *BARRINGTONIA RACEMOSA* FRUIT AND SEED EXTRACTS. Bol Latinoam Caribe Plantas Med Aromat 2004;3:88–92.
 84. Ahmed AAM, Ahmed SM. Extracts of Leaves and Seeds of the Neem Tree, *Azadirachta Indica*, As Environment-Oriented Molluscicides for Combating Schistosomiasis. African Freshwater Malacology, 2003, p. 235–49.
 85. Diallo D, Marston A, Terreux C, Toure Y, Smested Paulsen B, Hostettmann K. Screening of Malian Medicinal Plants for Antifungal, Larvicidal, Molluscicidal, Antioxidant and Radical Scavenging Activities. Phytotherapy Research 2001;15:401–6. <https://doi.org/10.1080/14786419.2014.931393>.
 86. Wanyonyi AW, Chhabra SC, Mkoji G, Njue W, Tarus PK. Molluscicidal and antimicrobial activity of *Solanum aculeastrum*. vol. 74. 2003. [https://doi.org/10.1016/S0367-326X\(03\)00030-3](https://doi.org/10.1016/S0367-326X(03)00030-3).
 87. Al-Zanbagi NA, Barrett J, Banaja AAA. Laboratory evaluation of the molluscicidal properties of some Saudi Arabian euphorbiales against *Biomphalaria pfeifferi*. Acta Trop 2001;78:23–9. [https://doi.org/10.1016/S0001-706X\(00\)00166-2](https://doi.org/10.1016/S0001-706X(00)00166-2).
 88. Adewoyin FB, Adewunmi CO, Omisore NO, Olorunmola FO, Elusiyan CA, Agvedahunsi JM. Larvicidal, molluscicidal and antitrichomonal effects of methanolic extract of *Lycopersicon esculentum* mill leaf from southwest Nigeria. Nigerian Journal of Natural Products and Medicines 2012;16:29–32.
 89. Agboola OI, Ajayi GO, Adesegun SA, Adesanya SA, Leavesò PÒ, Stemò A, et al. Comparative molluscicidal activities of fruit pericarp, leaves, seed and stem bark of *Blighia Unijugata baker*. Pharmacognosy Journal 2011;3:63–6. <https://doi.org/10.5530/pj.2011.25.11>.
 90. Mata RCS, de Mendonca DIMD, Vieira L, Dos Santos AF, da Silva LA, Gaspar JF, et al. Molluscicidal Activity of Compounds Isolated from *Euphorbia conspicua* N. E. Br. Journal of Brazilian Chemical Society 2011;22:1880–7.
 91. Changbunjong T, Wongwit W, Leemingsawat S, Tongtokit Y, Deesin V. Effect of crude extract of *Solanum xanthocarpum* against snails and mosquito larvae. Southeast Asian Journal of Tropical Medicine and Public Health 2010;41:320–5.
 92. Baloch IB, Baloch MK, Baloch AK. Schistosomiasis suppressing deoxyphorbol esters from *euphorbia cauducifolia* L. Latex. Planta Med 2010;76:809–14. <https://doi.org/10.1055/s-0029-1240691>.

93. Medina JM, Peixoto JLB, Silva AA, Haraguchi SK, Falavigna DLM, Zamuner MLM, et al. Evaluation of the molluscicidal and *Schistosoma mansoni* cercariae activity of *Croton floribundus* extracts and kaurenoic acid. *Revista Brasileira de Farmacognosia* 2009;19:207–11. <https://doi.org/10.1590/S0102-695X2009000200005>.
94. Abouzid S, Orihara Y, Kawanaka M. Molluscicidal activity of polyacetylenes from *Ambrosia maritima* hairy roots. *Nat Prod Commun* 2007;2:177–80. <https://doi.org/10.1177/1934578x0700200214>.
95. Batista JJ, de Araújo HDA, Aguiar TW de A, Ferreira SA de O, Lima M de V, Pereira DR, et al. Toxic, cytotoxic and genotoxic effect of saline extract and fraction of *Parkia pendula* seeds in the developmental stages of *Biomphalaria glabrata* (Say 1818 – intermediate host) and cercaricide activity against the infectious agent of schistosomiasis. *Acta Trop* 2022;228. <https://doi.org/10.1016/j.actatropica.2022.106312>.
96. Pereira LPLA, Ribeiro ECG, Brito MCA, Araruna FOS, Araruna FB, Leite JAC, et al. Molluscicidal and cercaricidal activities of the essential oil of *Dysphania ambrosioides* (L.) Mosyakin & Clemants: Implications for the control of schistosomiasis. *Acta Trop* 2022;230. <https://doi.org/10.1016/j.actatropica.2022.106393>.
97. Silva YRR, Silva LD, Rocha TL, Dos Santos DB, Bezerra JCB, Machado KB, et al. Molluscicidal activity of *Persea americana* mill. (lauraceae) stem bark ethanolic extract against the snail *Biomphalaria glabrata* (say, 1818): A novel plant-derived molluscicide? *An Acad Bras Cienc* 2020;92:1–16. <https://doi.org/10.1590/0001-376520202000715>.
98. Barros Gomes PR, Batista Reis J, Fernandes RP, Mouchrek Filho VE, De Souza AG, Fontenele MA, et al. Toxicity and molluscicidal activity of the essential oil *Pimenta dioica* against the snail *Biomphalaria glabrata*. *Rev Peru Biol* 2019;26:102–8.
99. Barros GRP, Oliveira MB, Sousa DA De, Caetano J, Fernandes RP, Louzeiro HC, et al. Larvicidal activity , molluscicide and toxicity of the essential oil of *Citrus limon* peels against , respectively , *Aedes aegypti* , *Biomphalaria glabrata* and *Artemia salina*. *Ecletica Quimica Journal* 2019;44:85–95. <https://doi.org/10.26850/1678-4618eqj.v44.4.p85-95>.
100. Faria RX, Rocha LM, Souza EPBSS, Almeida FB, Fernandes CP, Santos JAA. Molluscicidal activity of *Manilkara subsericea* (Mart.) dubard on *Biomphalaria glabrata* (Say, 1818). *Acta Trop* 2018;178:163–8. <https://doi.org/10.1016/j.actatropica.2017.11.012>.
101. Pereira LPLA, Dias CN, Miranda MiV, Firmo W da CA, Rosa C dos S, Brito MCA, et al. Molluscicidal effect of *Euphorbia umbellata* (Pax) Bruyns latex on *Biomphalaria glabrata*, *Schistosoma mansoni* host snail. *Journal of the Sao Paulo Institute of Tropical Medicine* 2017;40:1290–3. <https://doi.org/10.15537/smj.2019.12.24643>.
102. Sá JLF, Siqueira WN, Silva HAMF, Santos ML de O, Santos FTJ dos, Silva LRS, et al. Evaluation of molluscicidal activity of *Anadenanthera colubrina* extracts on adult mollusc and embryos of the species *Biomphalaria glabrata* (Say, 1818). *Scientia Plena* 2016;12:2–10. <https://doi.org/10.14808/sci.plena.2016.101001>.
- 103.[103] Rocha-filho CAA, Albuquerque LP, Silva LRS, Silva PCB, Coelho LCBB, Navarro DMAF, et al. Chemosphere Assessment of toxicity of *Moringa oleifera* flower extract to *Biomphalaria glabrata* , *Schistosoma mansoni* and *Artemia salina*. *Chemosphere* 2015;132:188–92. <https://doi.org/10.1016/j.chemosphere.2015.03.041>.
104. Feitosa A, Fonseca SA, César FA. A penta-substituted pyridine alkaloid from the rhizome of *Jatropha elliptica* (Pohl) Muell . Arg . is active against *Schistosoma mansoni* and *Biomphalaria glabrata*. *Parasitol Res* 2014;113:1077–84. <https://doi.org/10.1007/s00436-013-3743-2>.
105. Santos CCS, Araújo SS, Santos ALLM, Almeida ECV, Dias AS, Damascena NP, et al. Evaluation of the toxicity and molluscicidal and larvicidal activities of *schinopsis brasiliensis* stem bark extract and its fractions. *Revista Brasileira de Farmacognosia* 2014;24:298–303. <https://doi.org/10.1016/j.bjp.2014.07.006>.
106. Pereira Filho AA, França CRC, Oliveira D da S, Mendes RJ de A, Gonçalves J de RS, Rosa IG. Evaluation of the molluscicidal potential of hydroalcoholic extracts of *Jatropha gossypifolia* Linnaeus, 1753 on *Biomphalaria glabrata*. *Rev Inst Med Trop Sao Paulo* 2014;56:505–10. <https://doi.org/10.1590/S0036-46652014000600009>.
107. Coelho da Silva CLPA, Vargas TS, Baptista DF. Molluscicidal activity of *Moringa oleifera* on *Biomphalaria glabrata*: Integrated dynamics to the control of the snail host of *Schistosoma mansoni*. *Revista Brasileira de Farmacognosia* 2013;23:848–50. <https://doi.org/10.1590/S0102-695X2013000500019>.
108. Orechio P, Victor DM, Rapado LN, Sa A De, Fokoue HH, Scotti MT, et al. Schistosomiasis Control Using Piplartine against *Biomphalaria glabrata* at Different Developmental Stages. *Neglected Tropical Diseases* 2013;7:1–8. <https://doi.org/10.1371/journal.pntd.0002251>.
109. Rodrigues KADF, Dias CN, Do Amaral FMM, Moraes DFC, Mouchrek Filho VE, Andrade EHA, et al. Molluscicidal and larvicidal activities and essential oil composition of *Cymbopogon winterianus*. *Pharm Biol* 2013;51:1293–7. <https://doi.org/10.3109/13880209.2013.789536>.
110. Fontes-jr UR, Ramos CS, Serafini MR, Cavalcanti SCH, Alves PB, Lima GM, et al. Evaluation of the lethality of *Porophyllum ruderale* essential oil against *Biomphalaria glabrata* , *Aedes aegypti* and *Artemia salina*. *African Journal of Biological Science* 2012;11:3169–72. <https://doi.org/10.5897/AJB11.1314>.
111. Lima MG, Augusto RDC, Vasconcellos MC De. Metabolic changes in *Biomphalaria glabrata* infected with *Schistosoma mansoni* exposed to latex of *Euphorbia milii* solution versus times of preparation. *J Nat Prod* 2012;5:222–32.
112. Oliveira-filho EC, Geraldino BR, Coelho DR, De-carvalho RR, Paumgarten FJR. Comparative toxicity of *Euphorbia milii* latex and synthetic molluscicides to *Biomphalaria glabrata* embryos. *Chemosphere* 2010;81:218–27. <https://doi.org/10.1016/j.chemosphere.2010.06.038>.
113. Teles T V., Bonfim RR, Alves PB, Blank AF, Jesus HCR, Quintans-Jr LJ, et al. Composition and evaluation of the lethality of *Lippia gracilis* essential oil to adults of *Biomphalaria glabrata* and larvae of *Artemia salina*. *Afr J*

- Biotechnol 2010;9:8800–4. <https://doi.org/10.5897/AJB10.113>.
114. Santos NC, Dias CN, Coutinho-Moraes DF, Vilanova CM, Goncalves JDR, Souza N da S, et al. Toxicity and evaluation of molluscicidal activity of leaves of *Turnera ulmifolia* L. *Brazilian Journal of Bioscience* 2010;8:324–9.
115. Silva TMS, Camara CA, Freire KRL, Da Silva TG, Agra MDF, Bhattacharyya J. Steroidal glycoalkaloids and molluscicidal activity of *Solanum asperum* Rich. fruits. *J Braz Chem Soc* 2008;19:1048–52. <https://doi.org/10.1590/S0103-50532008000500033>.
116. Coutinho DF, Dias CS, Barbosa-Filho JM, Agra MF, Martins RM, Silva TMS, et al. Composition and molluscicidal activity of the essential oil from the stem bark of *ocotea bracteosa* (meisn.) mez. *Journal of Essential Oil Research* 2007;19:482–4. <https://doi.org/10.1080/10412905.2007.9699958>.
117. Mello-Silva CC, de Vasconcellos MC, Pinheiro J, de Azevedo Rodrigues M de L. Physiological Changes in *Biomphalaria glabrata* Say, 1818 (Pulmonata: Planorbidae) caused by sub-lethal concentrations on the latex of *Euphorbia splendens* var. *hislopianae* N.E.B (Euphorbiaceae). *Mem Inst Oswaldo Cruz* 2006;101:3–8. https://doi.org/10.5005/jp/books/12167_1.
118. Silva TMS, Câmara CA, Agra M de F, de Carvalho MG, Frana MT, Brandoline SVPB, et al. Molluscicidal activity of *Solanum* species of the Northeast of Brazil on *Biomphalaria glabrata*. vol. 77. 2006. <https://doi.org/10.1016/j.fitote.2006.05.007>.
119. Silva TMS, Batista MM, Camara CA, Agra MF. Molluscicidal activity of some Brazilian *Solanum* spp. (Solanaceae) against *Biomphalaria glabrata*. *Ann Trop Med Parasitol* 2005;99:419–25. <https://doi.org/10.1179/136485905X36208>.
120. Bezerra JCB, Silva IA, Ferreira HD, Ferri PH, Santos SC. Molluscicidal activity against *Biomphalaria glabrata* of Brazilian Cerrado medicinal plants. *Fitoterapia* 2002;73:428–30. [https://doi.org/10.1016/S0367-326X\(02\)00121-1](https://doi.org/10.1016/S0367-326X(02)00121-1).
121. Meléndez PA, Capriles VA. Molluscicidal activity of plants from Puerto Rico. *Ann Trop Med Parasitol* 2002;96:209–18. <https://doi.org/10.1179/000349802125000600>.
122. Dos Santos AF, Sant'Ana AEG. Molluscicidal properties of some species of *Annona*. *Phytomedicine* 2001;8:115–20. <https://doi.org/10.1078/0944-7113-00008>.
123. Schall VT, Vasconcellos MC, Rocha RS, Souza CP, Mendes NM. The control of the schistosome-transmitting snail *Biomphalaria glabrata* by the plant Molluscicide *Euphorbia splendens* var. *hislopianae* (syn *milli* Des. Moul): a longitudinal field study in an endemic area in Brazil. *Acta Trop* 2001;79:165–70.
124. Al-obaidi MJ, Abbas AH, Al-sa AJR, Al-husseiny IA. Schistosomiasis vector control using *Cucumis melo* Plant extracts with bioassay experiment 2018;18:425–30.
125. Al-abidi MJ, Abbad AH, Khanon AY, Ibrahim K. Using the *Thymus vulgaris* extracts to control the snail vector in Schistosomiasis (Part II). *The Iraqi Journal of Agricultural Sciences* 2018;49:111–23.
126. Al-Obaidi MJL, Abbasp AH, Al-Azzaup AAM, Waalp A. Survival rates of *Bulinus truncatus* as a way to determine the molluscicidal activity of *Ricinus Communis* extracts. *Iraqi Journal of Biotechnology* 2015;14:32–53.
127. El Babili F, Fabre N, Moulis C, Fouraste I. Molluscicidal activity against *Bulinus truncatus* of *Croton campestris*. vol. 77. 2006. <https://doi.org/10.1016/j.fitote.2006.03.003>.
128. Hannedouche S, Souchard JP, Jacquemond-Collet I, Moulis C. Molluscicidal and radical scavenging activity of quinones from the root bark of *Caryopteris x clandonensis*. vol. 73. 2002. [https://doi.org/10.1016/S0367-326X\(02\)00163-6](https://doi.org/10.1016/S0367-326X(02)00163-6).
129. Patel A V, Wright D, Blunden G, Sumner S, Rice J. Stable Molluscicide Formulation of an Aqueous Extract of *Euphorbia myrsinites*. *Phytotherapy Research* 2011;25:1412–4.
130. Apers S, Baronikova S, Sindambiwe JB, Witvrouw M, De Clercq E, Vanden Berghe D, et al. Antiviral, haemolytic and molluscicidal activities of triterpenoid saponins from *Maesa lanceolata*: Establishment of structure-activity relationships. *Planta Med* 2001;67:528–32. <https://doi.org/10.1055/s-2001-16489>.
131. Rug M, Ruppel A. Toxic activities of the plant *Jatropha curcas* against intermediate snail hosts and larvae of schistosomes. *Tropical Medicine and International Health* 2000;5:423–30. <https://doi.org/10.1046/j.1365-3156.2000.00573.x>.
132. Aminou HA, Mossalem HS, Alam-eldin Y. *Azadirachta indica* extracts on *Bulinus truncatus* snail and aquatic stages of *Schistosoma haematobium*. *Parasitologists United Journal* 2020;151–7. <https://doi.org/10.21608/puj.2020.39631.1085>.
133. Bakry FA, El-Garhy M, Abd El-Atti M, Th Atwa M. Effects of the Extracts of *Euphorbia Pulcherima* and *Atriplex Nummularia* on the Infectivity of *Schistosoma Haematobium* to *Bulinus Truncates* Snails. *Advances in Biomechanics* 2017;1:34–41. <https://doi.org/10.11648/j.abm.20170102.12>.
134. Hasheesh WS, Mohamed RT, El-Monem SA. Biological and physiological parameters of *Bulinus truncatus* snails exposed to methanol extract of the plant *Sesbania sesban* plant. *Adv Biol Chem* 2011;01:65–73. <https://doi.org/10.4236/abc.2011.13009>.
135. Okeke, O.C. and Ubachukwu PO. Molluscicidal Effects of *Talinum triangulare* on *Bulinus truncatus*. *Niger J Biotechnol* 2011;22:13–6.
136. Bakry FA, Mohamed RT, Hasheesh WS. Impact of methanol extract of *Adenium obesum* plant on some biochemical and biological parameters of *Bulinus truncatus* snails. *J Evol Biol Res* 2011;3:87–94.
137. Larhsini M, Sebbane R, Kchakech H, Markouk M, Bekkouche K, Abbad A, et al. Screening of some Moroccan plant extracts for molluscicidal activity. *Asian Journal of Experimental and Biological Sciences* 2010;1:964–7.
138. Lahlou M, Berrada R. Potential of essential oils in schistosomiasis control in Morocco. *International Journal of Aromatherapy* 2001;11:87–96. [https://doi.org/10.1016/S0962-4562\(01\)80023-0](https://doi.org/10.1016/S0962-4562(01)80023-0).
139. Hmamouchi M, Lahlou M, Agoumi A. Molluscicidal activity of some Moroccan medicinal plants. *Fitoterapia* 2000;71:308–14. [https://doi.org/10.1016/S0367-326X\(99\)00152-5](https://doi.org/10.1016/S0367-326X(99)00152-5).

140. Suleiman J, Singh K, Bala AY, Muhammad MT, Abdullahi A. Comparison for the Effectiveness of Column Purified Fractions of *Allium cepa* Bulbs and *Allium sativum* Cloves against *Bulinus globosus* (Intermediate Host of Comparison for the Effectiveness of Column Purified Fractions of *Allium cepa* Bulbs and *Allium sati*. *Asian Journal of Advanced Research and Reports* 2018;2:1–6. <https://doi.org/10.9734/ajarr/2018/v2i429774>.
141. Angaye TCN, Bassey SE, Ohimain EI, Izah SC, Asaigbe PI. Molluscicidal and synergicidal activities of the leaves of four Niger Delta Mangrove plants against schistosomiasis vectors. *Journal of Environmental Treatment Techniques* 2015;3:35–40.
142. Angaye TCN, Ohimain EI, Zige D V, Didi B, Biobelemoye N. Biocidal activities of Solvent Extracts of *Azadirachta indica* against Some Endemic Tropical Vector-borne Diseases. *International Journal of Tropical Disease* 2014;4:1198–208. <https://doi.org/10.9734/IJTDH/2014/11746>.
143. Okete J. Efficacy of the ethanolic extracts of *Talinum triangulare* for control of the fresh water snail, *Bulinus globosus*, the vector of urinary schistosomiasis. *International Journal of Pure and Applied Zoology* 2015;3:76–86.
144. Daniel UN, Onyemekara NN, Ohaletu UN, Nnoli MC. Schistosomiasis snail-intermediate host control, using a natural plant. *World J Pharm Pharm Sci* 2013;2:52–63.
145. Salawu OT, Odaibo AB. The molluscicidal effects of *Hyptis suaveolens* on different stages of *Bulinus globosus* in the laboratory. *Afr J Biotechnol* 2011;10:10241–7. <https://doi.org/10.5897/AJB10.415>.
146. Olofintoye LK. Comparative evaluation of Molluscicidal effects of *Securidaca longepedunculata* (Fres.) and *Tephrosia bracteolata* (Guilland Perr) on *Bulinus globosus*. *Journal of Parasitology and Vector Biology* 2010;2:37–41. <https://doi.org/10.21608/eajbsf.2010.17449>.
147. Jia T-W, Wang W, Sun L-P, Lv S, Kun Y, Zhang N-M, et al. Molluscicidal effectiveness of Luo-Wei, a novel plant-derived molluscicide, against *Oncomelania hupensis*, *biomphalaria alexandrina* and *Bulinus truncatus*. *Infect Dis Poverty* 2019;27:70–9.
148. Ke W, Tu C, Cao D, Lin X, Sun Q, Zhang Q. Molluscicidal activity and physiological toxicity of quaternary benzo [c] phenanthridine alkaloids (QBAs) from *Macleaya cordata* fruits on *Oncomelania hupensis*. *Neglected Tropical Diseases* 2019:1–16.
149. Ludevese FL, Ludevese-Pascual G, Portugaliza HP. Molluscicidal activity of *Jatropha curcas* L . crude leaf extracts against *Oncomelania hypensis quadrasii* Mollendorf. *J Sci Eng Technol* 2018;186:177–86.
150. Ke W, Lin X, Yu Z, Sun Q, Zhang Q. Molluscicidal activity and physiological toxicity of *Macleaya cordata* alkaloids components on snail *Oncomelania hupensis*. *Pestic Biochem Physiol* 2017;143:111–5. <https://doi.org/10.1016/j.pestbp.2017.08.016>.
151. Han BX, Chen J. Acacetin-7-rutinoside from *buddleja lindleyana*, A new molluscicidal agent against *oncomelania hupensis*. *Zeitschrift Fur Naturforschung - Section C Journal of Biosciences* 2014;69 C:186–90. <https://doi.org/10.5560/ZNC.2013-0179>.
152. Yang F, Long E, Wen J, Cao L, Zhu C, Hu H, et al. Linalool , derived from *Cinnamomum camphora* (L .) Presl leaf extracts , possesses molluscicidal activity against *Oncomelania hupensis* and inhibits infection of *Schistosoma japonicum*. *Parasit Vectors* 2014;7:1–13.
153. Li X, Shan X, Pan J, Yu P, Mao Z. Effects of the molluscicidal agent GA-C 13: 0 , a natural occurring ginkgolic acid , on snail mitochondria. *Pestic Biochem Physiol* 2012;103:115–20. <https://doi.org/10.1016/j.pestbp.2012.04.006>.
154. Chen YQ, Xu QM, Liu YL, Li XR, Yang SL, Zhuge HX. Laboratory evaluation of the molluscicidal activity of *pulsatilla Chinensis* (Bunge) regel saponins against the snail *oncomelania hupensis*. *Biomedical and Environmental Sciences* 2012;25:224–9. <https://doi.org/10.3967/0895-3988.2012.02.015>.
155. Sun Q, Yang Y, Zhang Q. Effect and mechanism of *Sapium sebiferum* against snail (*Oncomelania hupensis*) and control of schistosomiasis. *International Conference on Biomedical Engineering and Biotechnology* 2012:190–3. <https://doi.org/10.1109/iCBEB.2012.165>.
156. [156] Ming Z, Gui-Yin L, Jian-Guo Z, Li Z, Ke-Long H, Jin-Ming S, et al. Evaluation of molluscicidal activities of benzo[c]phenanthridine alkaloids from *Macleaya cordata* (Willd) R. Br. on snail hosts of *Schistosoma japonicum*. *Journal of Medicinal Plants Research* 2011;5:521–6.
157. Peng F, Liu M, Huang Q, Liu N, Yang H, Sun H, et al. Molluscicidal Effect of *Eomecon chionantha* alkaloids against *Oncomelania hupensis* snails. *Southeast Asian Journal of Tropical Medicine and Public Health* 2011;42:289–96.
158. Li Y, Chen J, Han B, Pan G, Guo D. Molluscicidal activities of *Ginkgo biloba* leaf against the snail *Oncomelania hupensis*. *Journal of Medicinal Plants Research* 2010;4:2466–72. <https://doi.org/10.5897/jmpr10.185>.
159. Han BX, Chen J, Yang X, Wang S, Li CG, Han FA. Molluscicidal activities of medicinal plants from eastern China against *Oncomelania hupensis*, the intermediate host of *Schistosoma japonicum*. *Brazilian Journal of Pharmacognosy* 2010;20:712–7. <https://doi.org/10.1590/S0102-695X2010005000018>.
160. Zou FC, Duan G, Xie YJ, Zhou Y, Dong GD, Lin RQ, et al. Molluscicidal activity of the plant *Eupatorium adenophorum* against *Oncomelania hupensis*, the intermediate host snail of *Schistosoma japonicum*. *Ann Trop Med Parasitol* 2009;103:549–53. <https://doi.org/10.1179/136485909X451780>.
161. Yang X ming, Chen S xia, Xia L, Chen J. Molluscicidal activity against *Oncomelania hupensis* of *Ginkgo biloba*. *Fitoterapia* 2008;79:250–4. <https://doi.org/10.1016/j.fitote.2007.11.030>.
162. Ke W, Yang J, Meng Z, Ma A. Evaluation of molluscicidal activities of *Arisaema tubers* extracts on the snail *Oncomelania hupensis*. *Pestic Biochem Physiol* 2008;92:129–32. <https://doi.org/10.1016/j.pestbp.2008.07.003>.
163. Chen SX, Wu L, Yang XM, Jiang XG, Li LG, Zhang RX, et al. Comparative molluscicidal action of extract of *Ginkgo biloba sarcotesta*, *arecoline* and *niclosamide* on snail hosts of *Schistosoma japonicum*. *Pestic Biochem Physiol* 2007;89:237–41. <https://doi.org/10.1016/j.pestbp.2007.07.010>.

164. Wei FH, Xu XJ, Liu JB, Dai YH, Dussart G, Trigwell J. Toxicology of a potential molluscicide derived from the plant *Solanum xanthocarpum*: A preliminary study. *Ann Trop Med Parasitol* 2002;96:325–31. <https://doi.org/10.1179/000349802125000727>.
165. Rug M, Ruppel A. Toxic activities of the plant *Jatropha curcas* against intermediate snail hosts and larvae of schistosomes. *Tropical Medicine and International Health* 2000;5:423–30. <https://doi.org/10.1046/j.1365-3156.2000.00573.x>.
166. Sukumaran D, Parashar BD, Rao KM. Evaluation of some plant molluscicides against a freshwater snail *Lymnaea luteola*, the vector of animal schistosomiasis evaluation of some plant molluscicides against a freshwater snail *Lymnaea luteola*, the vector of animal schistosomiasis. *Pharm Biol* 2002;40:450–5. <https://doi.org/10.1076/phbi.40.6.450.8448>.
167. Samah MA-K, Samah NE, Ismail ShA. Molluscicidal effects of acetone and ethanol extracts of clove (*Syzygium aromaticum*) against *Monacha cartusiana* (Gastropoda: Hygromiidae) snails under laboratory and field conditions at Sharkia Governorate. *Egyptian Journal of Plant Protection Research Institute* 2020;3:595–603.
168. El-Atti MA, A.A E, A.M. K, Elgohary WS. Control of the glassy clove snails *Monacha cartusiana* using Zingiber officinale extract as an ecofriendly molluscicide Mahmoud. *African Journal of Biological Science* 2019;15:101–15.
169. Abdel-mogib M, Dawidar AEM, Mortada MM, Raghieb HM. against *Monacha cartusiana* molluscicidal activity of *Balanites aegyptiaca* against *Monacha cartusiana*. *Pharm Biol* 2012;50:1326–9. <https://doi.org/10.3109/13880209.2012.674950>.
170. Abdelhady Ahmed EA. EFFECT OF CASTOR PLANT (*RICINUS COMMUNIS*) EXTRACTS ON THE TERRESTRIAL SNAIL *MONACHA OBSTRUCTA* (*FERUSSAC*). *Plant Arch* 2020;20:1734–42.
171. Hussien Ali IH, Mohammed El-Sayed SA, Ali Fathey R, Abd El-Halim SM. Molluscicidal and antifeedant impacts of some weed extracts on the land snail *Monacha obstructa* (Pfeiffer). *Fayoum Journal of Agricultural Research and Development* 2020;34:204–17.
172. Alzabib AA, Abobakr Y, Al-sarar AS, Hussein HI, Basudan OA, El-gamal AA, et al. Molluscicidal activity of cardiac glycosides isolated from *Adenium obesum*. *Pest Manag Sci* 2019;75:2770–5. <https://doi.org/10.1002/ps.5388>.
173. Preet S. Laboratory evaluation of Molluscicidal and Cercaricidal Potential of *Artemisia annua* (Family: Asteraceae). *Ann Biol Res* 2010;2:373–83.
174. Muraleedharan K, Murthy SHK. Preliminary screening of leaves of *Leucas aspera* and *Parthenium hysterophorus* against fresh water snail, *Indoplanorbis exustus*. *Indian Journal of Veterinary Research* 2009;17:8–12.
175. Al-sarar A, Hussein H, Abobakr Y, Bayoumi A. Molluscicidal Activity of Methomyl and Cardenolide Extracts from *Calotropis procera* and *Adenium arabicum* Against the Land Snail *Monacha cantiana*. *Molecules* 2012;17:5310–8. <https://doi.org/10.3390/molecules17055310>.
176. Usman AM, Shinkafi SA. Molluscicidal Activity of Some Selected Plants on Freshwater Snail *Lanistes Ovum*. *International Journal of Science for Global Sustainability* 2016;2:24–37.
177. Ali TH, Abdul Jabbar Hamed A. The Molluscicidal Activity of *Melia azadirachta* on The Fresh water snail *Physa acuta* (*Draparnaud* , 1805) The Molluscicidal Activity of *Melia azadirachta* on The Fresh water snail *Physa acuta* (*Draparnaud* , 1805). *Journal of Pure and Applied Sciences* 2019;5:1–7.
178. Dib R, Makhoul K, Maalouf R. Preliminary bioactivity investigation of *Styrax officinalis* fruit extract as potential biopesticide. *Journal of Pharmacognosy and Phytotherapy* 2016;8:209–13. <https://doi.org/10.5897/JPP2016.0422>.
179. Souza BA De, Cristina L, Chicarino ED, Cristina E, Bessa DA. Phytochemical and Molluscicidal activity of *Mikania glomerata* Sprengel (*Asteraceae*) in Different Lifestages of *Subulina octona* (*Mollusca* , *Subulinidae*). *Brazilian Archives of Biology and Technology* 2014;57:261–8.
180. Souza BA, Silva LC Da, Chicarino ED, Bessa ECA. Preliminary phytochemical screening and molluscicidal activity of the aqueous extract of *Bidens pilosa* Linné (*Asteraceae*) in *Subulina octona* (*Mollusca* , *Subulinidae*). *Annals of the Brazilian Academy of Sciences* 2013;85:1557–66.
181. Akinpelu BA, A. DC, I. AF, O. IE, O.O. O. Effect of stem - bark of *Erythrophleum suaveolens* (Guill. Perri.) saponin on fresh water snail (*Lanistes lybicus*) tissues. *Afr J Environ Sci Tech* 2012;6:446–51. <https://doi.org/10.5897/ajest12.007>.
182. Silva L, Durço E, Pinheiro J, Bessa ECDA. Effect of *Solanum paniculatum* leaf extract on food consumption , fertility and carbohydrate metabolism of *Bradybaena similaris* snail . *J Nat Prod* 2013;6:168–76.
183. Kariuki ST, Kariuki JM, Mailu BM, Muchiri DR. *Phytolacca octandra* (L.), *Phytolacca dodecandra* (LHerit) and *Balanites aegyptiaca* (L.) extracts as potential molluscicides of schistosomiasis transmitting snails. *Journal of Medicinal Plants Research* 2016;10:823–8. <https://doi.org/10.5897/jmpr2016.6237>.
184. Nnamdi UC, Chuka EP, Chukwudi UK, Chinelo U, Okwudiri IM. Molluscicidal Assessment of Aqueous Extract of *Moringa oleifera* Lam seed on *Bulinus* Snail for the control of Schistosomiasis Molluscicidal Assessment of Aqueous Extract of *Moringa oleifera* Lam Seed on *Bulinus* Snail for the Control of Schistosomiasis. *South Asian Journal of Research in Microbiology* 2020;7:31–9. <https://doi.org/10.9734/SAJRM/2020>.
185. Mtemeli FL, Walter I, Shoko R. Molluscicidal effects of pumpkin seed extracts on *Schistosoma* vectors 2020:1–13.
186. Eisa AI, Mohammed Elfaki TE, Abdallah MA, Elfaki M. Molluscicidal Activity of *Khaya Grandifoliola* Extracts Against Freshwater Snails in Khartoum State- Sudan. *International Journal of Academic Health and Medical Research* 2019;3:1–4.
187. Kindiki MG. Molluscicidal Activity of Selected Plant Extracts. 2014.
188. Caixeta SC, Magalhães LG, de Melo NI, L. Wakabayashi KA, Paula Aguiar G de, Paula Aguiar D de, et al. Chemical Composition and *in Vitro* Schistosomicidal Activity of the Essential Oil of *Plectranthus Neochilus* Grown in Southeast Brazil. *Chem Biodivers* 2011. <https://doi.org/10.1002/cbdv.201100167>.

189. Soares MH, Dias HJ, Vieira TM, Mendes Souza MG, Cruz AF, Badoco FR, et al. Chemical Composition, Antibacterial, Schistosomicidal, and Cytotoxic Activities of the Essential Oil of *Dysphania Ambrosioides* (L.) *Mosyakin* & *Clemants* (Chenopodiaceae). *Chem Biodivers* 2017. <https://doi.org/10.1002/cbdv.201700149>.
190. Domínguez-Vigil IG, Mata-Cárdenas BD, Esquivel-Ferriño PC, Ávalos-Alanís FG, Vargas-Villarreal J, Rayo Camacho-Corona M del. Antigiardial Activity of *Foeniculum Vulgare* Hexane Extract and Some of Its Constituents. *Plants* 2022. <https://doi.org/10.3390/plants11172212>.
191. Tripathi SM, Singh DK. Molluscicidal activity of *Punica granatum* bark and *Canna indica* root. *Brazilian Journal of Medical and Biological Research* 2000;33:1351–5. <https://doi.org/10.1590/S0100-879X2000001100014>.
192. Dossaji S, Kubo I. Quercetin 3-(2"-Galloylglucoside), A Molluscicidal Flavanoid from *Polygonum Senegalense*. *Phytochemistry* 1980;19:482.
193. Marston A, Hostettmann K. Plant molluscicides. *Phytochemistry* 1985;24:639–52. [https://doi.org/10.1016/S0031-9422\(00\)84870-0](https://doi.org/10.1016/S0031-9422(00)84870-0).
194. Doctor TR. Molluscicidal Performance of *Kae Farang* (*Gliciridia sepium*, JacqStrudel and Siam Weed (*Chromolaena odorata*, Linn.) Leaf Extracts on Golden Snail (*Pomacea caniculata*, Linn.) 2019.
195. Singh V, Kumar P, Singh VK, Singh DK. Effect of abiotic factors on the molluscicidal activity of oleoresin of *Zingiber officinale* against the snail *Lymnaea acuminata*. *Nat Sci (Irvine)* 2010;02:1148–54. <https://doi.org/10.4236/ns.2010.210142>.
196. Abubakar A, Bala AY, Singh K. Plant Molluscicides and their Modes of Action: a Review. *International Journal of Advanced Scientific Research* 2017;2:37–49.
197. Sharifi-Rad J, Sureda A, Tenore GC, Daglia M, Sharifi-Rad M, Valussi M, et al. Biological Activities of Essential Oils: From Plant Chemoecology to Traditional Healing Systems. *Molecules* 2017. <https://doi.org/10.3390/molecules22010070>.
198. Kalinda C, Chimbari MJ, Grant WE, Wang H, Odhiambo JN, Mukaratirwa S. Simulation of Population Dynamics of *Bulinus Globosus*: Effects of Environmental Temperature on Production of *Schistosoma Haematobium* Cercariae. *PLoS Negl Trop Dis* 2018. <https://doi.org/10.1371/journal.pntd.0006651>.
199. Henderson I, Triebkorn R. Chemical control of terrestrial gastropods. *Molluscs as Crop Pests* 2009:1–31. <https://doi.org/10.1079/9780851993201.0001>.
200. Hamed M, El-Amin S, Abdel-Fattah AS. Effect of some environmental conditions on the molluscicidal activity of *yucca filamentosa* “marginata” and *cestrum purpureum*. *Pharmacologyonline* 2015;2:142–7.